



Next-generation Close-kin Mark Recapture:

**Using SNPs to identify half- sibling pairs in Southern
Bluefin Tuna and estimate abundance, mortality and
selectivity**

**Campbell Davies, Mark Bravington, Paige Eveson, Matt Lansdell, Jordan Aulich and
Peter Grewe**

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Researcher Contact Details

Name: Campbell Davies
Address: CSIRO Marine Laboratories
Hobart, TAS 7030
Phone: 03 62325044
Fax:
Email: campbell.davies@csiro.au

FRDC Contact Details

Address: 25 Geils Court
Deakin ACT 2600
Phone: 02 6285 0400
Fax: 02 6285 0499
Email: frdc@frdc.com.au
Web: www.frdc.com.au

In submitting this report, the researcher has agreed to FRDC publishing this material in its edited form.

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Executive Summary

This report presents the results of the first application of Close-Kin Mark-Recapture (CKMR) using both Parent-Offspring Pairs (POP) and Half-sibling Pairs (HSP). This application to Southern Bluefin Tuna (SBT) has been successful, providing a decadal time series of absolute abundance, total mortality and selectivity of adults. The method and the results have been reviewed and accepted by the Scientific Committee of the Commission for the Conservation of Southern Bluefin Tuna (CCSBT) and CKMR is now adopted and funded by the CCSBT as a fisheries-independent method for monitoring the rebuilding of the spawner abundance. While the focus of this report is SBT, this extended method of CKMR (i.e. POP+HSP), developed by CSIRO, is applicable to most teleosts (bony fishes) and will have wide applicability to Australian and international fisheries.

The original driver for the development of the CKMR approach was the need to provide robust estimates of population status that were not subject to the often-unquantifiable statistical biases associated with commercial catch per unit effort (CPUE) data and, in the case of SBT, large unreported longline catches that undermined the conventional stock assessment. The results of that original application in 2012 provided the first direct, fishery-independent estimates of adult abundance. The incorporation of these POP data in the CCSBT stock assessment substantially reduced the uncertainty in estimates of stock status as well. The benefits of the method saw CSIRO, CCSBT and the Australian Government continue to invest in sample collection, strategic methods development and design studies for SBT, in particular, and CKMR more generally. This project is built on several important developments since the first CKMR project;

- i) New theory and methods on the use of more distant kin (i.e., half-siblings) developed by CSIRO, which provide the basis to estimate total adult mortality independently of selectivity;
- ii) Next-Generation-Sequencing method selected and refined by CSIRO that allow HSPs to be reliably identified at a cost that makes large-scale genotyping cost-effective for large populations such as SBT, and;
- iii) Accumulation of an extended time-series of SBT samples funded by CSIRO, DAWR and CCSBT.

Collectively, these developments provided the basis to estimate abundance and total mortality of spawning SBT (and to separate the confounding effect of selectivity) using methods and data that are free of the biases associated with fishery dependent CPUE data and the impacts of the historical unreported catches. The POP and HSP data series were reported to the CCSBT and incorporated into the conditioning of the CCSBT Operating Models, as part of the regular assessment of stock status in 2017. They are also being used in the development and testing of candidate Management Procedures (MP) for future setting of global catch levels.

In the context of this larger collaborative effort, the specific aims of this FRDC project were:

1. Process archived tissue samples, extract DNA, and genotype them (~16,000 individuals, 2006-14)
2. Combine those genotypes with those from related CCSBT project (~2000 more samples 2015-16).
3. Estimate time series of total adult abundance, spawning potential and total mortality for the spawning population, using the methods developed by CSIRO.
4. Report outcomes to SBTMAC, AFMA and CCSBT Scientific Committee for incorporation into the 2017 update of the CCSBT Operating Models.

Parent-Offspring and Half-sibling Pairs were identified using specifically designed software and SNP assays for SBT, developed by CSIRO with DArT Pty Ltd, and samples of adults and juveniles collected between 2006 and 2015. In total, DNA was extracted from ~ 17,000 individuals with a total of ~15,000 individuals (4,238 adults and 10,952 juveniles) analysed for

POP and HSP following DNA and genotyping quality control. A total of 76 POPs (including the same 45 found in the original CKMR study), 140 definite HSPs and 4 full-sibling pairs were identified. We are confident that false-positives are negligible, but we estimate that about 10% of true HSPs would fail to meet our cut-off criterion for HSP. This false-negative rate is built into the CKMR model that uses the kin-pairs. Examination of mitochondrial DNA indicated that about 65 of the 140 HSPs shared a mother, whereas 75 shared a father. This is consistent with an equal sex-ratio in the adult component of the SBT population.

The estimates of abundance from the new POP+HSP model and data were similar to the values from the 2012 POP-only study, with the new estimates of spawner abundance being about 10% higher on average. This degree of change is consistent with sampling variability. The overall summary statistics of biomass and numerical abundance varied relatively little across the set of alternative model options explored, but there were differences in the age-specific components of the adult population (e.g., numbers of 8 year-olds recruiting to the adult population). The models with estimated, rather than fixed, selectivity predict more old and fewer young adult fish. All options explored show strong incoming cohorts of 8 year-olds from about 2012 onwards and, by 2014, those cohorts have started to make an impact on overall spawner abundance. Given the strength of these incoming cohorts, substantial upward trends in spawner abundance would be expected from 2015 onwards as these recent recruits to the adult component of the population continue to grow.

A key difference from the original CKMR SBT study is that the HSPs data provide a direct signal on total adult mortality. The estimates from the new method are broadly consistent with the overall mortality inferred under the assumptions of the original POP-only model. However, the new model does show some preference for a somewhat higher survival for young adults, and an overall dome-shaped selectivity. This difference would have some effect on turnover rates of the adult component of the population, and estimated abundance of incoming 8-year-old recruitments. It will be possible to examine this effect in more detail in the future as the time series of POPs and HSP increases.

With the addition of HSPs, it becomes possible to check two assumptions that previously had to be made for the original POP-only analysis: that selectivity was strictly proportional to residence-time in the Indonesian fishery, and that there would be a low proportion of siblings (half or full) within each year's sample of juveniles. The new data do validate the latter assumption and show that the first appears to be a reasonable approximation; moreover, with the addition of HSPs, it becomes possible to avoid making that selectivity assumption at all.

Future periodic updates of the CKMR models should be used to review the evidence in the new data and continue to test the validity of this assumption.

The 76 POPs, 140 HSPs, and the estimated false-negative rate for HSP were included in the preliminary reconditioning of the CCSBT Operating Models (OMs) in preparation for a full stock assessment in 2017¹. The inclusion of the CKMR data and parameter (false-negative rate) has substantially improved the fits of the CCSBT OMs and reduced the overall structural uncertainty across the grid of operating models, including natural mortality for adults². The results were reviewed by the Operating Model and Management Procedure Technical Group and the Extended

¹ Hillary, R., Preece, A. and Davies, C. (2017b). Summary of initial incorporation of the Half-Sibling Pair (HSP) data in the CCSBT Operating Model. Working Paper to CCSBT-OMMP Webinar, 21 July 2017. Commission for the Conservation of Southern Bluefin Tuna.

Hillary, R.M., Preece, A.L., Davies, C.R., Takahashi, N., Sakai, O. and Itoh, T. (2017c). Reconditioning of the CCSBT Operating Model in 2017. Working paper prepared for the Extended Scientific Committee for the Twenty-Second Meeting of the Scientific Committee, Yogyakarta, Indonesia, 28 August – 2 September 2017. CCSBT-ESC/1709/14.

² Anon (2017). Report of the OMMP Technical Webinar on the incorporation of half-sibling pairs in the CCSBT OMs for the 2017 update of stock status. Working paper CCSBT-ESC/0817/36 to the Twenty-Second Meeting of the Extended Scientific Committee. Yogyakarta, Indonesia, 28 August – 2 September 2017.

https://www.ccsbt.org/en/system/files/ESC22_36_report_2017HSPWebMeeting_0.pdf

Anon (2017b). Report of the Twenty-Second Meeting of the Extended Scientific Committee. Yogyakarta, Indonesia, 28 August – 2 September 2017.

https://www.ccsbt.org/sites/default/files/userfiles/file/docs_english/meetings/meeting_reports/ccsbt_24/report_of_SC22.pdf.

Scientific Committee at their 2017 meetings. The use of the data, and associated modifications to the CCSBT OMs to accommodate them, were accepted by the CCSBT for use in the 2017 stock assessment. In addition, the empirical POP and HSP data series were accepted as input monitoring series for use in the development and testing of new Candidate Management Procedures. The support for the approach by CCSBT also includes ongoing funding of tissue sampling and kin identification (POP and HSP) for use in stock assessment, management procedures and direct monitoring of adult abundance as part of the CCSBT rebuilding plan for the stock.

The outputs from this and the related projects have SBT-specific and more general implications for monitoring and management of fisheries. In the case of SBT, it has demonstrated the power and utility of the POP+HSP method of CKMR to estimate three of the central parameters for fisheries stock assessment: abundance of adults, total mortality and average age of the adult component of the SBT stock (and, thereby, selectivity). This has been done in a stand-alone CKMR assessment framework that is independent of the catch and effort data for the major longline fisheries. All this has substantially reduced the uncertainty in the state of the stock, improved the stability of the CCSBT OMs used for stock assessment and, as a direct result, substantially increased national and international confidence in the estimates of stock status. In addition, the time-series of POP and HSPs have been shown to be highly informative when used in Candidate Management Procedures (MPs) designed to set the level of global catches of SBT. As a result, the Australian Government, stakeholders and the CCSBT have agreed to the use of these CKMR data series (POPs and/or HSP) in Candidate MPs being developed and tested in the CCSBT Management Strategy Evaluation process. A final MP will be selected by the CCSBT to replace the current MP and used to set the TAC for 2020 and beyond.

Importantly, this science has been developed and delivered in a manner that has engaged both national stakeholders and managers and members of the international management organisation, which has resulted in substantive “buy-in” and ongoing support. This has direct ongoing benefits for the Australian Government, industry and wider stakeholders, as well as for the other members of the CCSBT. These include the quality, utility and cost of the information being used to monitor the stock and set global catch levels, and the confidence in and acceptance of the decisions based on the scientific advice.

In the wider fisheries stock assessment and management context, this project has delivered the first application of the what is likely to become the “template” method of CKMR for most teleost populations. The strong non-linear increase in fecundity with size/age in most teleosts means it will be necessary to use the POP+HSP method to generate unbiased estimates of adult abundance, and to avoid having to rely on assumptions about CPUE or selectivity. The shift from microsatellite (used in the original study) to the specific SNP assays developed with DArT Pty Ltd for SBT, combined with streamlining of laboratory work-flows by, has demonstrated it is possible to efficiently process the large sample sizes (17,000 individuals in ~ 6 months) and identify HSP sufficiently accurately to provide the required samples sizes of POPs and HSPs for precise estimates of abundance and mortality. These empirical results are consistent with those expected from CKMR theory and, combined with positive results from CKMR studies for other species, continues to demonstrate the breadth of utility and cost-effectiveness of the approach for fisheries and conservation management.

The impact of this work for SBT and other species would be enhanced through:

1. Continued refinement of lab workflows to reduce cost, time and increase quality control.
2. Refinement of software and genotyping approaches to increase resolution for identifying HSP and more distant kin.
3. Ensuring long-term viability and consistency of adult (Indonesia) and juvenile (Australian) sampling programs.
4. Building quantitative capacity for the wider application of the method to Australian fisheries.

Keywords

Southern Bluefin Tuna, close-kin mark-recapture, stock assessment, fisheries independent, abundance estimation, mortality, selectivity, management procedure, regional fisheries management organisation, CCSBT.

Introduction

Background

Abundance, mortality and selectivity are three of the most influential parameters in fisheries stock assessment and management; however, they are commonly the most difficult to estimate with the requisite level of accuracy and precision. In most fisheries, these important parameters are estimated using a time series of relative abundance (e.g. CPUE from commercial catch and effort statistics) and associated catch composition data, which are “integrated” in a statistical catch at age, or length, model. This approach, while appropriate for some data-rich fisheries, has substantial limitations for a range of fisheries that either: do not have the requisite time-series of data required, and/or the available time series are subject to substantial, and largely unquantifiable, biases.

Original application of Close-Kin Mark-Recapture to Southern Bluefin Tuna

Close-Kin Mark-Recapture (CKMR) is a suite of methods to estimate the abundance of adults and other important demographic parameters using information on the frequency of closely related individuals (i.e. kin) in samples (Bravington et al, 2016a). The first large-scale application was for Southern Bluefin Tuna (SBT) (Bravington et al, 2014; 2016b) where it was developed as an abundance estimator independent of commercial catch per unit effort (CPUE) and total catch data.

The impetus to do so for SBT was three-fold:

1. There was no direct index of abundance for the spawning stock, i.e. the mature component of the population and the primary target of the rebuilding plan for the stock. Instead, it was extrapolated from sub-adult abundance estimates via a suite of integrated stock assessment models (CCSBT Operating Models (CCSBT OMs));
2. There were (and are) unresolved issues associated with statistical methods and interpretation of longline CPUE as an index of abundance of the harvested age classes of SBT (Davies et al, 2008; Anon. 2014) and
3. In addition to 2 above, there were revelations of large, long-term, unreported catches from the longline fisheries that generated unquantifiable uncertainty (Anon. 2006, Polacheck 2012) in the stock status, to the extent that the Extended Scientific Committee (ESC) of the CCSBT could no longer conduct a stock assessment in the conventional sense (Anon 2006). The last point, in particular, increased the urgency for developing more reliable sources of abundance information for the spawning stock, which is the primary focus of the CCSBT rebuilding plan.

The initial SBT application of Close-kin Mark-Recapture (CKMR) used specifically designed microsatellite loci (Bravington et al, 2016) to identify 45 Parent-Offspring-Pairs (POPs) in about 14,000 samples of known spawning adults (Indonesia) and known-age juveniles (Great Australian Bight). These were embedded in a statistical mark-recapture framework and, combined into a stand-alone mini-assessment of the adult component of the population that used length and age composition data from Indonesian longline catches on the spawning ground, plus histological information on relative daily fecundity-at-size (Farley et al, 2014). This stand-alone assessment was able to estimate a time-series of absolute spawning stock biomass, effective annual fecundity-at-size and total mortality rate of the mature component of the population. Full details of the sampling design, genotyping, quality control, procedures for identifying POPs, estimation model, and independent review process are provided in (Bravington et al, 2014). The approach and the final results were reviewed by the CCSBT Extended Scientific Committee in 2012 and 2013 and accepted as: (i) a valid fishery-

independent estimate of spawning stock abundance for SBT, and (ii) as valid input data (i.e. the 45 POP information and associated comparisons) for the CCSBT OMs (Hillary et al, 2012, 2013; Anon, 2013).

Parent-Offspring Pairs in CCSBT Operating Models

The CCSBT Operating Models (OM) are a set (n=432 in 2017) of integrated statistical-catch-at-age models used for development and testing of Management Procedures (MP) (Hillary et al, 2015) and periodic assessments of stock status (e.g. Preece et al 2014, Hillary et al, 2017). This large number of models is used so as both plausible ranges of uncertainty in status and dynamics of the stock is encapsulated in assessments of stock status and MP testing. The unquantifiable uncertainty resulting from the unreported catches means that a variety of historical-catch scenarios, provided by the CCSBT Commission, are used to scale the standardised CPUE from the reported catch and effort data from the primary longline fleet (Anon 2009, 2014). This CPUE of largely juvenile and sub-adult age classes was (and is) the primary abundance index used in the CCSBT OMs. The reason for having a set of models, rather than just one, is to accommodate different scenarios about historical catch and other structural uncertainties in the dynamic of the stocks and fisheries. Other sources of abundance information include conventional tagging data from the 1990s, and a relative abundance index of juveniles from a scientific aerial survey from 1993-2014 (e.g. Eveson et al, 2012). The close-kin data (i.e. the outcome of each juvenile-adult comparisons to identify POPs) are incorporated into the OMs directly as mark-recapture data, with a corresponding mark-recapture component in the likelihood (Hillary et al, 2012, 2013). Two substantive adjustments to the OMs were required to make it structurally compatible with the CKMR data: first to deal with the absence of sex- and length-substructure in the OM (as CKMR fundamentally requires that both be accounted for somehow); and, second, to change the form of the maturity ogive from knife-edge to logistic, consistent with the results on fecundity-at-size from the stand-alone CKMR analysis (Bravington et al, 2012; Hillary et al, 2012; Anon. 2013).

The CKMR data were very informative when incorporated into the OMs (Anon. 2013). This in part reflected the new absolute abundance information on the spawning component of the population the CKMR data provided, where there previously was no information; however, it was also because some of the adult cohorts in the close-kin data were also observed as juveniles in the 1990s conventional tagging data. Since both data sets contain information on abundance and mortality, the combination of the two data sets constrain the plausible fits and parameter space considerably. This resulted in the exclusion of the more pessimistic OM scenarios and a revision to the OM “grid” (Hillary et al, 2013; Anon. 2013)

Beyond Parent-Offspring-Pairs and microsatellites

The potential of CKMR for directly estimating absolute abundance and other key demographic parameters for natural resource management, has led to substantial investments in the theory and practice subsequent to the first tranche of SBT-related work. This has included:

1. Development of demographic CKMR models that can use Half-Sibling Pairs (HSPs): where two animals have one shared parent, as well as Parent-Offspring Pairs (Bravington et al, 2016a).
2. Reviewing and testing the suitability and cost-effectiveness of different Next Generation Sequencing platforms (e.g. DArT, RadSeq, Sequenom, and GBS) for large-scale close-kin genotyping to find HSPs and POPs (See Grewe and Davies in related projects).
3. Development of a general statistical/demographical theory for CKMR (Bravington et al, 2016a)

4. Design and implementation of CKMR studies for other species (especially sharks) with very different sampling and demography (e.g. where only juveniles can be sampled) (e.g. Hillary et al, 2018)
5. Design work for CKMR as a long-term monitoring tool for CCSBT, using HSPs as well as POPs (Bravington and Davies, 2013; Bravington, 2014 and Bravington et al, 2015). The long-term use of CKMR, along with gene-tagging to estimate recruitment (Preece et al, 2015), is now endorsed by the CCSBT Scientific Committee and funded under the CCSBT Scientific Research Program (Anon 2018).

In addition to the fundamental development work referred to above, the results reported here draw directly on investments and outputs from three related projects:

1. *Long-term monitoring of Indonesian catches from the SBT spawning grounds – CSIRO-Agency for Marine and Fisheries Research, Indonesia*

Landings of SBT from Indonesian vessels fishing on the spawning grounds south of Bali-Java have been monitored since 1993, with otoliths collected from 1994, as part of a collaborative program between the research agency of the Ministry for Marine Affairs and Fisheries of Indonesia (Farley et al, 2014; Farley et al, 2015; Farley et al, 2017). The sampling program was extended in 2006 to include tissue samples, providing the adult samples for the original CKMR study, and has been maintained through support from CSIRO, MMAF-Indonesia and DAWR and, now, CCSBT (see below).

2. *Collection and genotyping of 2015-16 SBT samples for close-kin – CCSBT-CSIRO*

Since 2014, this project has supported the annual collection of tissue samples of adult SBT from Benoa, Indonesia, and 3-year-old juveniles from Port Lincoln, Australia, as well as DNA extraction, archiving and sequencing of the DNA (Farley et al, 2018).

3. *Estimating abundance, mortality and selectivity using Close-kin pairs - CSIRO.*

This project developed the SNP markers, assays and genotyping pipelines required to accurately and reliably identify Half-Sibling and Parent-Offspring pairs from samples of SBT DNA and the modelling framework to estimate spawning abundance, total mortality and selectivity using these two types of close-kin data (Bravington et al, 2017). This modelling framework and analysis pipelines have been used in this project, along with the POPs and HSP identified from the genotyping, to generate a time series of absolute adult abundance, mortality and selectivity for SBT from 2002 through to 2014.

Need

As noted, the original driver for the development of the CKMR approach was to provide robust estimates of population status that were not subject to the statistical biases associated with commercial CPUE data and, in the case of SBT, large unreported longline catches that undermine the conventional assessment. The current project extends the application of the CKMR approach for SBT by using a new method, which incorporates both Parent-Offspring Pairs and Half-Sibling Pairs (Bravington et al, 2015), and approximately 15,000 samples of SBT spanning 2006-14, with an additional 2,000 samples supported by the related CCSBT-CSIRO project. Specifically, this FRDC project supports processing, DNA extraction and sequencing by NGS of archived samples of SBT from 2006-14 and the estimation of the time series of spawning biomass from 2002-14 using Parent-Offspring and Half-Sibling Pairs that is independent of CPUE and total catch data.

In doing so, the outputs of this project (and the related projects) directly address three major needs for the monitoring, assessment and management of SBT:

1. Reducing the uncertainty in the status of the spawning stock and the impact of unreported longline catches on the assessment results.

- i) Output: A stand-alone CKMR assessment of the abundance, total mortality and selectivity of the spawning stock that is independent of longline CPUE and catch data.
 - ii) Output: A time series of Parent-Offspring and Half-sibling Pairs (and associated comparisons) as an input to the CCSBT Operating Models, as part of the 2017 CCSBT assessment of stock status.
2. A direct index of the spawning stock for use in a Management Procedure.
 - i) The same time series of Parent-Offspring and Half-sibling Pairs can be used to generate two independent empirical indices of the spawning stock (one based on POP and a second from HSP) that can be used as inputs to Candidate Management Procedures as part of developing a new CCSBT MP for recommending the global TAC from 2020.
3. Cost-effective, repeatable methods for long-term monitoring.
 - i) The primary motivation for the development of the SNP assays was to reduce the cost of processing and reliance on human expertise for “scoring” the microsatellite markers used in the original study. The shift to SNP markers, which are effectively the base unit of DNA, is important in future proofing the approach, substantially reducing the time and cost, and development of standardised protocols and analysis pipelines.

Objectives

1. Process archived tissue samples, extract DNA and genotype (~16,000 individuals, 2006-14³)
2. Combine genotypes from 1 with those from related CCSBT project (2015-16)
3. Estimate time series of total adult abundance, spawning potential and total mortality for the spawning population
4. Report outcomes to SBTMAC, AFMA and CCSBT Scientific Committee for incorporation into 2017 update of the CCSBT Operating Model.

³ Note, that the precise years processed under each project changed from those originally proposed due to poor DNA quality associated with the 2015 adult samples and the timing of availability of the data from the 2016 samples. Notwithstanding these necessary changes, approximately 16,000 individuals were analysed as proposed. See Methods for full details.

Methods

From tissue to kin

Sample collection, tissue processing, DNA extraction and archiving

Sampling of adults takes place in Benoa, Indonesia, during processing of catches from the longline fishery on the spawning ground, with tissue and otoliths collected at the same time. Sampling of juveniles caught by the Australian the purse seine fishery takes place in Port Lincoln, Australia. Sampling is done in the processing factories during harvest from grow out pens, some 3-6 months after capture. Samples consist of a biopsy containing ~300mg of tissue, which are placed in 2.0 mL cryovials, frozen, and transported to the CSIRO Marine Laboratories in Hobart.

Tissues are held at -80°C until sub-sampled in preparation for DNA extraction. For each fish selected for subsampling, a ~15mg slice of tissue is weighed and placed into an extraction chamber for tissue digestion. An Eppendorf EP motion robot completes the DNA extraction and produces two final 96-well plates: a sequencing plate, and a replica DNA archive plate. Each plate contains DNA from 92 individuals, as well as two blanks and two control tissue samples; the position of which allow unique identification of each plate for quality control (QC) purposes. The archive plates are stored frozen at -80°C where they remain unless required for further testing.

DNA sequencing

The sequencing plates are sent for sequencing at Diversity Array Technologies Pty Ltd (DART <https://www.diversityarrays.com>), Canberra, using a specific variant of Genotyping- By-Sequencing designed by CSIRO and DART for close-kin purposes, known as DARTcap. This involves laboratory pre-processing of the plates; analysis using a high-throughput sequencer and assays for a specific set of ~1500 SNP loci; and bioinformatic analysis of the terabytes of resulting data, to produce specific data summaries for each fish at each SNP locus of interest.

Genotyping for kin identification

The final step prior to kin identification takes the data summaries provided by DART and turns them into multi-locus genotypes for each individual fish i.e., for each fish and each locus, the pair of alleles inferred to be present. This genotype-calling entails some quite complicated algorithms developed at CSIRO specifically for DARTcap sequencing data, and also estimates the genotyping error-rates for each locus. The latter is essential for robust identification of kin, in particular HSP, and associated uncertainty.

Kin Identification

The final step prior to the CKMR modelling itself is kin-finding, which is based on the inferred genotypes and the error-rates from the multi-locus genotyping. For kin identification CSIRO has developed generic algorithms (i.e. not specific to DARTcap) from basic statistical principles (For those interested in more detail, these are summarized in the Appendix and in section 5 of Bravington et al, 2016b). Control of rates of false-positive and false-negative kin is crucial to this process, since ~100,000,000 comparisons might be needed to find only ~100 true kin-pairs.

Maternal and Paternal Half-sibling Pairs

It is possible to determine whether each HSP is the result of sharing the same father (Paternal HSP) or mother (Maternal HSP) using genetic analysis of mitochondrial DNA of samples of identified HSPs. The comparison of maternal and paternal HSP provides insights into differences in how fecundity varies with age between males and females, and on the true sex ratio of adults. The mtDNA data are incorporated directly in the standalone CKMR estimation model (see below).

The CCSBT OMs are not sex structured and thus cannot use the maternal/paternal information in the HSPs; instead, it is assumed that there is little difference among adult male and females both in total numbers, and in how age and length affect individual fecundity. Thus, the standalone CKMR model, which does separate adults by sex and treats the MHSPs and PHSPs separately, can provide a test of the CCSBT OM assumptions.

Large-scale processing of archived tissue samples for DNA extraction began in October 2016, following in principle agreement between CSIRO and FRDC on funding arrangements. All DNA extractions were complete by January 2017 and sequencing of ~ 16,000 fish at DArT began in February 2017; the full set of sequencing- files were received by CSIRO at the end of March 2017. In parallel, CSIRO developed quality control (QC), genotype-calling and kin-finding algorithms suitable for the new type of genetic data. From April to June, these algorithms were refined and applied to deliver reliable sets of POPs and HSPs (Bravington, 2017) suitable for use in the 2017 reconditioning of the CCSBT OMs (Hillary et al, 2017b).

Table 1: Summary of DNA extractions and samples successfully sequenced for SNPs by DArTcap for identification of Parent-Offspring-Pairs and Half-Sibling-Pairs funded by FRDC 2016-044.

Year/season	Source	Sampled	DNA Extracted	Sequenced
2006	Port Lincoln	4042	1472	1468
2007	Port Lincoln	4085	1472	1443
2008	Port Lincoln	4138	1564	1488
2009	Port Lincoln	4100	1473	1458
2010	Port Lincoln	4071	1472	1467
2011	Port Lincoln	4000	1012	1011
2012	Port Lincoln	4000	1012	1000
2013	Port Lincoln	1600	1012	998
2014	Port Lincoln	1600	1012	998
2005-06	Indonesia	216	0	0
2006-07	Indonesia	1520	0	0
2007-08	Indonesia	1594	0	0
2008-09	Indonesia	1637	0	0
2010-11	Indonesia	1013	1012	1011
2011-12	Indonesia	565	552	549
2012-13	Indonesia	1381	1012	998
2013-14	Indonesia	1642	1011	991
Total		41204	15088	14880

Table 2: Summary of DNA extractions and samples successfully sequenced for SNPs by DArTcap for identification of Parent-Offspring-Pairs and Half-Sibling-Pairs funded by related CCSBT project. * Note the original proposal was to include the 2014-15 Indonesian samples. However, these were not included due to tissue quality issues with this particular year. The 2009-10 samples were substituted in their place.

Year/season	Source	Sampled	DNA Extracted	Sequenced
2015	Port Lincoln	1600	1011	1005
2009-10	Indonesia	1200	1012	1012
2014-15*	Indonesia	1609	0	0
Total		4409	2023	2017

Table 3: Summary of final number of samples genotyped and used in kin-finding by location and year

Year/season	Indonesia (Adults)	Port Lincoln (Juveniles)
2006	0	1281
2007	0	1305
2008	0	1315
2009	0	1317
2010	943	1284
2011	931	938
2012	527	844
2013	933	873
2014	904	873
2015	0	922
Total	4238	10952

Incorporation of Parent-Offspring and Half-Sibling Pairs in CCSBT Operating Models for 2017 stock assessment

The Parent-Offspring-Pair data were available in time to be incorporated into the OM updates for June 2017 meeting of the Operating Model and Management Procedure (OMMP) Technical Group in preparation for the 2017 assessment of stock status (Hillary et al, 2017a). The HSPs were reported separately (Bravington, 2017). There was insufficient time to incorporate these data into the CCSBT OMs prior to OMMP8, due to the more complex nature of the analysis and quality control procedures, relative to the POPs. Notwithstanding this, the OMMP Technical Group, both data sets were available for the OMMP meeting to review, and the technical group recommended the new POP and HSP data be included in the 2017 assessment, conditional on a review of the results of incorporation of the HSP data (Anon, 2017a). The HSP data were incorporated into the OMs following the June meeting (Hillary et al, 2017b) and the results reviewed at a special webinar meeting of the OMMP Technical Group. The OMMP was satisfied with the results and agreed to include the HSP data series in the reference set of OMs for the 2017 stock assessment, and to an additional sensitivity run to examine the implications of a specific assumption relating to the HSP (Anon, 2017b).

Overview of Stand-alone CKMR model with POPs and HSPs

There are four main changes from the stand-alone CKMR model used in the original application to SBT (Bravington et al, 2014; 2016a). These are:

1. Extending the time-series of data through to 2014 (length/sex/age frequencies; genotypes);
2. Modifying the length-frequency model to allow for annual changes in selectivity;
3. Inclusion of Half-Sibling Pairs
4. “Freeing” selectivity from fecundity.

There are also some minor differences associated with the different nature of the input data and providing greater consistency with assumptions of the CCSBT Operating Models, where it reasonable to do so (e.g. plus-group at age 30, rather than 25).

The most important change is the inclusion of HSPs. The key benefit of HSPs is that they lead rather directly to an estimate of average adult total mortality (z), or more accurately of the

rate-of-turnover of Total Reproductive Output⁴. Parent-Offspring Pairs alone do not carry intrinsic information on z . In the original application to SBT Bravington et al (2014) addressed this limitation by assuming:

- that selectivity was directly proportional to residence time on the spawning grounds;
- that male and female adult mortality rates were equal.

These assumptions could not be directly tested within the original framework, nor was there other information (e.g. electronic tagging data) with which to test it.

Incorporating HSPs into the log-likelihood of the stand-alone model is relatively straightforward. The underlying equations are unambiguous about how that should be done (see Appendix, and the explanation of HSP probabilities in section 3.9 of Bravington et al, 2016b). The greater complication lies in allowing a more flexible selectivity-at-length relationship, plus allowing that to vary from year to year.

There are four main datasets included in the stand-alone model, each of which contributes separately to the log-likelihood:

1. Parent-Offspring Pairs (POPs): The number of comparisons, and number of POPs found, broken down by year-of-adult-capture, adult sex, length and age, and year-of-juvenile birth. Note that comparisons are not made between adults caught in one season and juveniles born in that same season.
2. Half-Sibling Pairs (HSPs): The number of comparisons, and a number of Maternal HSPs (shared mother) and Paternal PHSPs (shared father), broken down by birth-years of the two juveniles being compared⁵. Note, comparisons are not made between juveniles from the same cohort.
3. Age, given Length and Sex (A@LS) in Indonesian otolith subsamples: That is, given the length and sex of a fish, what was the estimated age?
4. Length frequency composition of the monitored component of the catch from the spawning ground (LSfreq): Samples from Indonesian fishery that are selected for otolith extraction (though not all otoliths are subsequently read). These are assumed to constitute a random subsample of landings in Bali.

Note that the first three inputs are truly “fishery-independent”, in that they are driven only by fish biology and the qualitative circumstances of sampling, rather than by changes in fishing/fleet behaviour. There is a clear, albeit complicated, logical path for how to model POPs, HSPs, and A@LS statistically i.e., for how the demographic parameters set the statistical distributions of each. There is also no reason to expect these sub-models to change with time (except if there are cohort-specific growth changes).

However, the fourth dataset, LSfreq, is subject to the selectivity of the Indonesian longline fishery; partly as a result of fish behaviour, and partly via boats fishing in different regions and/or with different gear setups (e.g. longline setting depth). This could be modelled in many different ways. The sample sizes are much bigger than for the other three datasets, so unless the intrinsic variability in selectivity can be successfully allowed for, this data will tend to dominate the likelihood and distort the fit.

The original SBT model (Bravington et al, 2014) assumed that numbers-at-length-and-sex followed an over-dispersed multinomial distribution each year, with expected values predicted from the population dynamics and the estimated residence-time at length and sex. The extent of over-dispersion (corresponding to about an 8-fold effective reduction over the actual

⁴ Total Reproductive Output (TRO), in some arbitrary but fixed unit such as “equivalent average 16yo SBT”— basically the same as SSB, except that TRO is a more accurate measure of what SSB is “trying” to measure.

⁵ It is possible to determine whether a HSP is by maternal or paternal descent by examination of the mitochondrial DNA, which is inherited from the mother only. See section 3.3 of appendix of Bravington et al, 2017, for further detail.

sample size of LSfreq) was previously estimated based on empirical comparisons of length frequencies in adjacent years. The over-dispersion adjustment is applied to each centimetre length class separately; it is very simple, but fails to capture large-scale shifts in selectivity, whereby many adjacent length-classes may become over- or under-represented in a given year. Since 2010, such systematic shifts have been apparent in the length and age frequency data from the Indonesian spawning ground fishery (Farley et al, 2017), and over-dispersion alone is clearly no longer adequate to account for this shift. Instead, a more complex annually-variable, spline-based selectivity adjustment, estimated via random effects, has been incorporated (see Appendix for details). A similar approach is used in the CCSBT OMs, albeit age-based rather than length-and-sex-based.

Exploring key model assumptions

The addition of HSP data allows a much wider range of model options to be explored than was possible with POPs alone. The CKMR model has been developed in a statistical framework (Restricted or Residual Maximum Likelihood (REML)), which is based on an explicit log-likelihood. This allows different “options” of the model, e.g., if survival rate is allowed to depend on sex as well as age, to be explored seamlessly, with accompanying diagnostics, to identify a preferred option from the set explored.

We have examined a reasonable number of options (see Appendix for full details) and the results indicate that the main issues to consider are:

- **selectivity**: constrained to match residence-time, as per Bravington et al, 2014, or more flexible (e.g. allowing dome-shape).
- **α HSP**: estimated (by sex) or fixed at 1. This parameter is there to allow for any unexpected discrepancy between the observed numbers of POPs and of HSPs, which could arise if some adults are systematically under-represented in the HSPs. For example, adults who systematically tend to breed offspring that migrate to South Africa in summer, rather than to the Great Australian Bight. Ideally, we would like to see α HSP =1, but this needs to be checked because there may be some surprises in reproductive biology. It is logically impossible to have α HSP >1 (see Appendix for details), although an estimate might come out slightly higher than 1 just by chance.
- **LSfreq weighting**: should the LSfreq be used at full strength, or down-weighted? If the selectivity model is adequate, then there should be no need to apply an overall down-weighting to the LSfreq; but if it is not, then an option with down-weighted LSfreq may be more robust.

Almost all options described here fit the CKMR data and the A@LS data quite well, but there remain some tensions with the LSfreq data, which warrant further investigation; in particular, no option can yet accurately reproduce the change in observed sex frequency.

Results and Discussion

Collection and Sample Processing

The first two objectives for the project related to the processing of archived samples from the original CKMR project for SBT and those collected subsequently by i) CSIRO and ii) CCSBT funded projects. The total number of samples from the respective projects are summarised in table 1 (Archived and new FRDC-CSIRO) and table 2 (CCSBT-CSIRO) and the final number of adult and juvenile samples sequenced and used in the analysis and modelling are given in table 3. In summary a total of 17,111 individuals were processed (DNA extracted and sequenced) and 15,190 individuals (4,238 adults and 10,952 juveniles) included in the final analysis following quality control of the sequencing data and resulting genotypes; effectively meeting objectives 1 and 2.

Kin Identification

Parent Off-spring Pairs

The microsatellites used in the first round of SBT CKMR were adequate for finding POPs using Mendelian-exclusion principles (see Appendix of Bravington 2014). However, a lot of statistical processing was required to control false-positive rates to an acceptable level and demonstrate that false-negatives must be rare. As the new DArTcap genotyping (Bravington et al, 2015) has been designed with the goal of identifying HSPs, which is much harder than finding POPs; finding POPs ought to be easier and clearer with the DArTcap genotyping results.

As in Bravington et al, 2014, we again identified POPs using a classification statistic based on Mendelian-exclusion, but some changes to the method were required to deal with the new features of DArTcap data (see Appendix, Bravington et al, 2017b). Figure 1 shows part of the histogram of the modified exclusion statistic, referred to as the Weighted-PSEUDO-EXCLUSION (WPSEX) statistic, across all genotyped adult-juvenile pairs (about 66,000,000 comparisons). The POPs are visible as a small bump on the LHS. Most of the entire histogram (to the right-hand side of the figure) has been left out, as otherwise the true POPs are too few to be visible compared to the very large peak of unrelated pairs. The peak of the unrelated pairs distribution is at 0.116, which is precisely where theory predicts it should be based on the allele frequencies of each locus.

The very large peak of unrelated pairs drops off very quickly to the left of ~ 0.08 , and the fattish tail around 0.055 to 0.075 will contain a number of adult/juvenile HSPs or GGP (Grandparent-Grandoff-spring Pairs), which should be somewhat rarer than true POPs on demographic grounds. The POPs are clearly separated from non-POPs. This separation is much more obvious with the new DArTcap data than it was for the original microsatellite data, demonstrating that the 1500 low-information SNP loci from DArTcap are performing better than 25 high-information microsatellite loci, at roughly half the cost.

The results presented in Figure 1 only uses only adults from 2010 onwards and, hence, exclude the POPs already found via microsatellites. However, we also DArTcapped those particular pairs-of-samples already identified as POPs in the original study, as a check, and they were all clearly identified as POPs using the new DArTcap method. Interestingly, we also processed one curious adult/juvenile pair from 2012, which was clearly not a POP according to microsatellites, but nevertheless remarkably close (just two unambiguous Mendelian exclusions in 25 loci compared). The DArTcap WPSEX statistic for this pair was around 0.06, consistent with being a Grandparent-Grandoff-spring Pair or HSP.

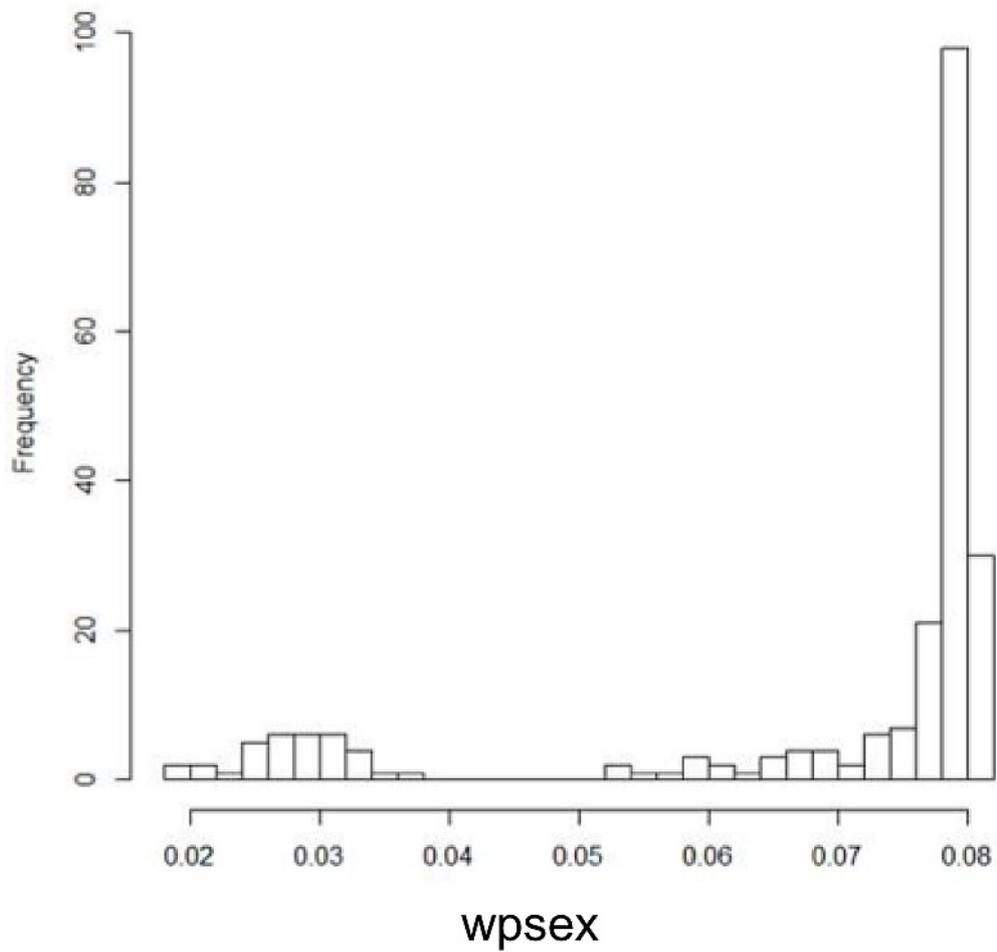


Figure 1: Identification of Parent-Offspring Pairs (POPs) via weighted-pseudo-exclusion (wpsex). Low values (left-hand side) indicate POPs. The x-axis has been truncated on the right-hand side to omit the majority of the very large peak of unrelated pairs, which would dwarf the small “bump” of POPs on the LHS if included.

Table 4: Distribution of Parent-Offspring Pairs by year juvenile birth year (rows) and adult Capture year (columns).

	2006	2007	2008	2009	2010	2011	2012	2013	2014
2002	0	0	0	0	0	0	0	0	0
2003	0	5	1	2	0	0	0	1	0
2004	0	2	0	0	3	0	0	0	0
2005	1	4	5	4	1	0	1	1	2
2006	0	4	3	2	0	0	0	0	0
2007	0	0	3	4	1	3	2	0	2
2008	0	0	0	0	0	1	1	1	0
2009	0	0	0	0	0	1	1	1	0
2010	0	0	0	0	0	3	1	4	0
2011	0	0	0	0	0	0	1	2	1
2012	0	0	0	0	0	0	0	1	1

The distribution of the POPs found in this study appear generally consistent with results from the previous study (Table 4). No POPs were found where the parent was caught in the same year as the offspring was born. As noted above, such comparisons are excluded from the model anyway, to avoid potential bias. As in 2012, we also read the ages of all adults in POPs using otoliths collected in Indonesia at the same time as genetic sampling. The modal age of parents at the time of offspring's birth was 13 or 14 years. All but one of the parents were 8yo or more at off-spring-birth, as was the case in the previous study. There was, however, one parent inferred to be 7-year-old: 14yo at capture, 7 years after its off-spring was born. The uncertainty of ± 1 year in otolith-derived ages is not uncommon, so this may be an age-estimation error.

The distribution of POPs in Bravington et al, 2014 indicated that adults younger than 12 years may spawn every second year, i.e. exhibiting skip-spawning behaviour. Only two of the new POPs in the current study involved an adult caught at age 12 or less, but in both cases the number of years between off-spring birth and adult capture was even. This result adds additional weight to the hypothesis that younger adult SBT skip-spawn.

Half-Sibling Pairs

Among 10,809 juvenile genotypes, we found 140 definite HSPs and 4 Full Sibling Pairs (FSPs). The true number of HSPs is expected to be about 10% higher than 140, because of false-negatives that are inevitable (and expected) consequence of the statistical criteria used to ensure exclusion of all false-positives (see Bravington et al, 2015). The HSPs and FSPs are quite clearly identified in Figure 2. The distributions of the Pseudo-Likelihood Ratio (PLOD, see Bravington et al 2015, Appendix C.2) test statistic match the predictions of the genetic theory, indicating that the new genotyping and HSP-finding processes work reliably. The details of the genotyping and HSP-finding analyses are provided in the Appendix.

The proportion of HSPs where both individuals were caught in the same year is somewhat higher than would be expected under a completely random breeding scenario⁶. This is

⁶ In a completely random breeding scenario, every juvenile sampled would have randomly selected its mother independently from the pool of potential mothers (weighted according to their relative fecundities), and likewise its father. The key word is "independently" (i.e. between juveniles in our sample). This does apply to juveniles in different cohorts, but the HSP data show that is not entirely true for juveniles in the same cohort.

evidence of “lucky litters”, i.e. variable survival between spawning events⁷ within each year class, which is also the only way to explain the 4 FSPs identified⁸. However, SBT is clearly not a sweepstake reproduction species; the proportion of juveniles in same-cohort HSPs is still very small (<1%). This result confirms the conclusion of the original CKMR analysis (Bravington et al, 2014) that it is a reasonable approximation to treat all POP comparisons as statistically independent. Note that within-cohort HSP comparisons are not used in our CKMR models; the HSP information comes entirely from cross-cohort comparisons.

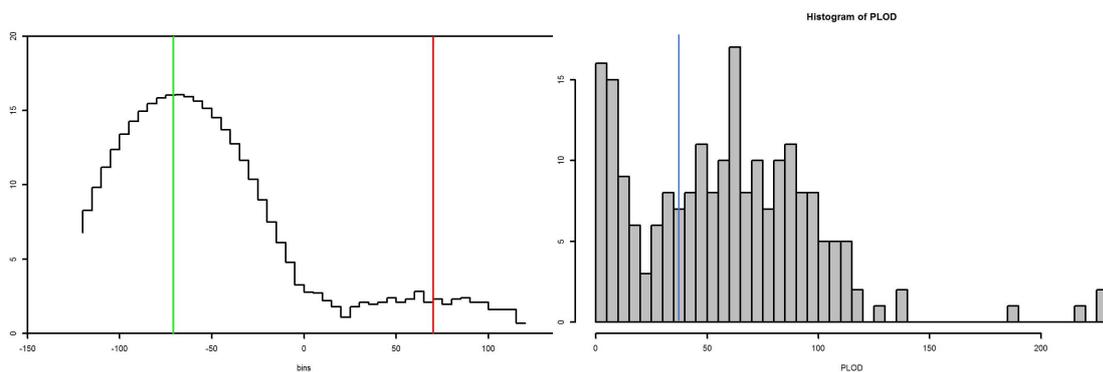


Figure 2: Half-Sibling Pairs (HSP). Left-hand side: log histogram (number of comparisons between individuals versus level of genetic relatedness (PLODs), with relatedness increasing to the right) to show all individual comparisons included in the analysis. The large dome on the left of zero is the Unrelated Pairs (UPs). The flatter, smaller dome on the right is the HSP and a few FSP. The green and red lines are theoretical means for the distribution of UPs and HSPs, respectively. Right-hand side panel: is a histogram of PLODs above zero, which shows the HSP (PLOD ~35-130) and full-sibling pairs (PLODs above ~140). The cut-off at PLOD=37 (blue vertical line) was chosen to exclude false-positive HSP and implies a false-negative rate of ~10% for true HSP with PLOD < 37.

Maternal/Paternal ratio

From the analysis of mitochondrial DNA, the Maternal/Paternal proportion in the HSPs is close to 50:50 (i.e. whether the shared parent is the Mother or the Father), both for same-cohort and cross-cohort HSPs. The mean number of cohorts separating each HSP is very similar for Maternal vs Paternal HSPs, so that the SSB turnover rate must be similar for both sexes, something which is not biologically obvious in advance. This validates an assumption underlying the exploratory use of combined-maternal-and-paternal HSPs in the CCSBT operating models for the 2017 assessment (Hillary et al, 2017b), where adult sexes are not distinguished.

⁷ Each SBT on the breeding ground spawns on many nights per year. Post-fertilization larval survival rates may well differ between nights.

⁸ The chance of a female breeding twice independently with the same male is inverse to adult abundance, so cross-cohort FSPs should be about a million times rarer than HSPs. The same applies to same-cohort FSPs, unless some spawning-events, where one female and a small number of courting males all release eggs and sperm together, have higher post-fertilization survival than others. Unsurprisingly, all 4 FSPs found were same-cohort.

Stand-alone CKMR assessment

Abundance and Spawning biomass- POP only versus POP+HSP models

The estimates of abundance from the new POP+HSP model and data are fairly similar to the values from the previous POP-only study (Bravington et al, 2014). Figure 3 shows that the new estimates of SSB are about 10% higher on average – a degree of change which is consistent with that expected from sampling variability, given there were 45 POPs available in the original study, whereas there are 76 in total in the updated data series. The downward trend in spawner abundance also seems milder with the addition of the new data. This general pattern of new estimates being slightly higher, and trends apparently weaker, is repeated in most of the other abundance-related statistics.

It is noteworthy that there is less of a difference between the old and new estimates earlier in the series, i.e. in 2002 relative to 2010. CKMR data tend to contain more information in the earlier part of a time series than the more recent part, because there have been more opportunities to recapture parents of juveniles born in the early years. Hence, any inference made in 2010 about abundance close to 2010 would have been particularly uncertain.

Similarly, inferences about 2014 made now are less certain than those about 2010 made now. All options suggest a small preponderance of females in the adult population, in the range 52–58%. The increase in fecundity-at-age for older (female) SBT appears less marked in the new data than was apparent in Bravington et al, 2014 (Figure 4). The original estimates were of course based on just 20 female parents in total.

Overall, there are no obvious strong differences between results from the original SBT study (using POP data only, up to 2010) and the new model (using HSPs as well as POPs, and data through to 2014).

Influence of Alternative Model assumptions on abundance related estimates

Turning to the new model options, the overall summary statistics of biomass and numerical abundance change rather little across the model options explored (Figure 5), but there are differences in the age-specific components (n_{16p} , n_{PLUS} , R_{cts}), whereby the options with estimated, rather than fixed, selectivity predict more old and fewer young fish.

All model options show very strong incoming cohorts of 8-year-olds (lower RHS panel) from about 2012 onwards, with the exception of the dotted-green option, where the LS_{freq} has been down-weighted by a factor of 100 so that it carries almost no information. Unsurprisingly, that option is slower to recognize incoming cohorts. By 2014, those cohorts have started to make an impact on overall SSB and TRO, so substantial upward trends in TRO and SSB would be expected from 2015 onwards as these recent adults grow bigger. These cohorts were born from 2004 onwards.

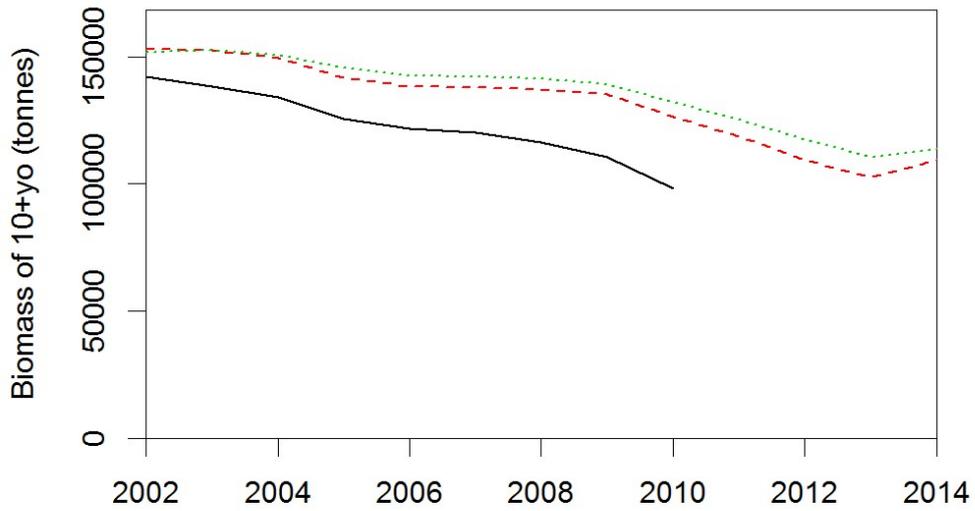


Figure 3: The original POP-only and new POP+HSP model estimates of adult biomass presented as total biomass of SBT aged 10 years and older. Black is the original model, which has been refitted using up-to-date software and modified code yielding very similar numbers to Bravington et al, 2014; red is the new POP+HSP model with fixed selectivity; green is new POP+HSP model with estimated selectivity. Similar ratios are seen for numbers of SBT aged 10 years and older.

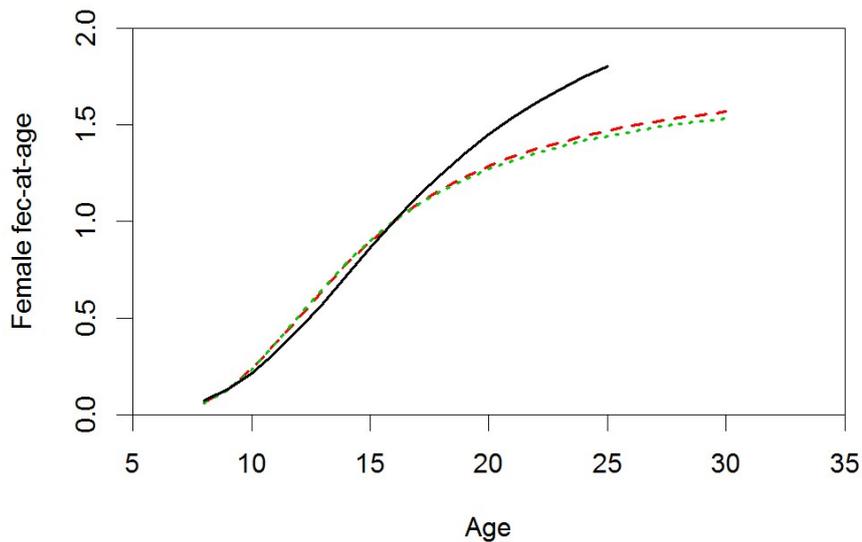


Figure 4: Female fecundity-at-age, normalised to 16 years old, from the original POP-only model (Bravington et al, 2014) and the new POP+HSP model and data. Black is original; red is new with fixed selectivity; green is new with estimated selectivity.

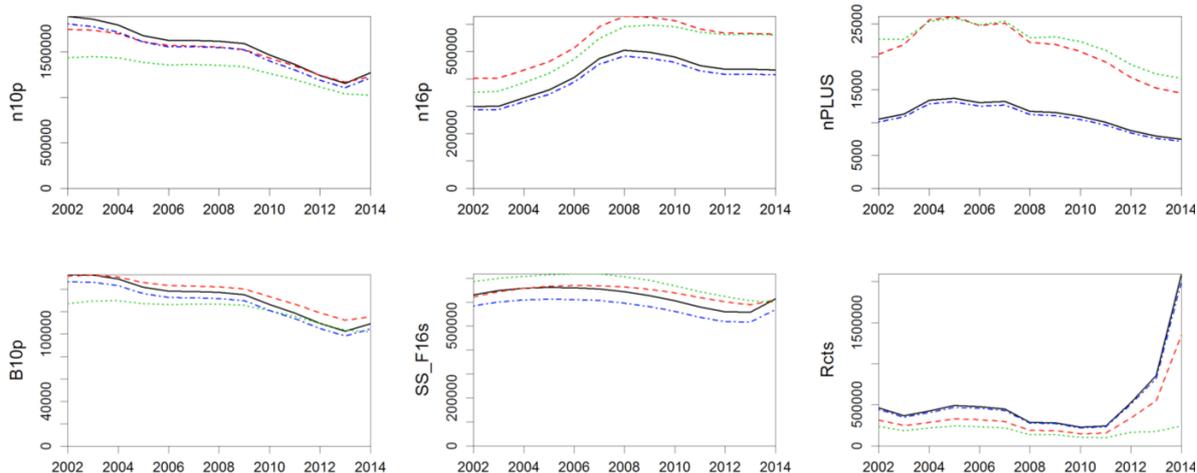


Figure 5: Abundance related estimates for six different components of the adult age-structure (each panel) from four different POP+HSP models (different lines). n10p = abundance of 10+ year-olds; n16p = abundance of 16+ year-olds, nPLUS = abundance of 30+ year-olds; Rcts = abundance of 8 year-olds recruiting to the adult component of the population; SS_F16s = Total Reproductive Output of females standardised to units of 16 year-old female; B10p = biomass of 10+ year-old. Lines reflect different models: black solid: fixed selectivity, with α_{HSP} fixed at 1; blue dot-dash: as for black except with α_{HSP} estimated; green dotted: as black except with LSfreq downweighted 100-fold; red dash: as black except with selectivity estimated.

Total Mortality and Selectivity

- Results from exploring the influence of a wide range of model options indicated the main differences in terms of selectivity and total mortality occur in the following areas:
- The estimated-selectivity options give somewhat higher survival for younger adults (0.85 vs 0.80), and preference for dome-shaped selectivity. The estimated mean age of 8+yos in the population is about 1 year greater for the estimated-selectivity model, with that gap increasing slightly over the time series (2002–14).
- A selection of estimated selectivity curves is shown in Figure 6. The preference for a dome-shaped selectivity persists even when the LSfreq is heavily down-weighted (right-hand column).
- When α_{HSP} and selectivity are both free, rather than fixed, and when LSfreq is not downweighted, the resulting estimate of α_{HSP} takes is nonsensical (α is estimated at ~ 2 , which is more than 3 standard errors away from its maximum mathematically-plausible value of 1). For this option, the biomass estimates do change substantially, but as the parameter estimates make no biological or mathematical sense, there is no reason to consider this option plausible.

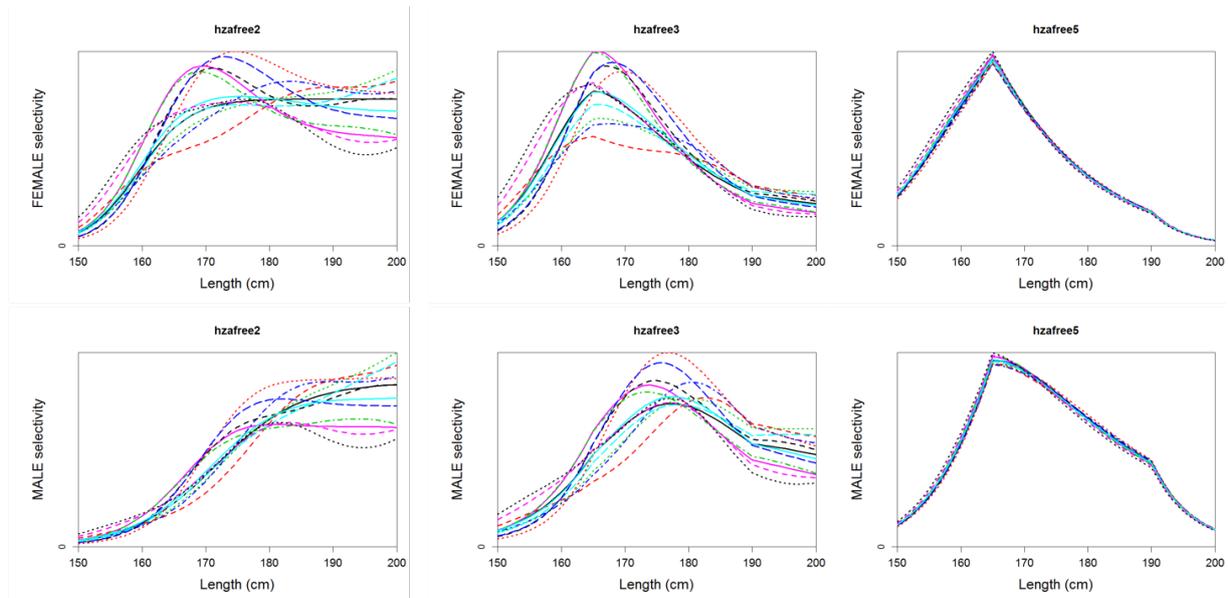


Figure 6: Annual selectivities-at-length for alternative models with females on the top row and males below. The left column are “fixed” selectivity (i.e. matching the residence time); the middle column is “free” (estimated) selectivity; the right column is free selectivity but with LSfreq downweighted by 100-fold. Different-coloured lines are for different years, showing annual variability. The abrupt kinks in the right-hand plots result from the piecewise-linear form used for base selectivity-at-length. These are smoothed out in the middle plots via the year-specific selectivity adjustments, but in the right-hand plots the LSfreq data is so heavily down-weighted that there is no incentive for the model to bother with much smoothing.

A few other options have been investigated (e.g. different z by sex; more flexible size-fecundity relationship for males), but without much obvious effect. Down-weighting the LSfreq data has interesting effects. The preference for dome-shaped selectivity remains, even though there is little impetus from the (down-weighted) data to drive it directly. The fit to the CKMR summary statistics is in fact slightly better with down-weighted LSfreq (e.g. less tension between HSPs and POPs when α HSP is fixed at 1), perhaps because the down-weighted options have more freedom to adjust other parameters, such as growth rates, when there is no need to match the LSfreq data closely.

Whether selectivity is fixed or free, the down-weighted options prefer higher survival rates for young adults (from 0.80 to 0.87 if selectivity is fixed; from 0.85 to 0.91 if selectivity is free). There is still remarkably little change in the main summary statistics (numbers of adults, SSB, TRO), but the higher survival rates do correspond to substantially different estimates of the absolute abundance of 8yo recruitments, and of overall age composition.

The internal CV on most biomass and numerical-abundance statistics is about 10–15%. It is systematically lower when α HSP is fixed, as, in this case, HSPs as well as POPs contribute directly to the absolute level of abundance, so the latter is being estimated based on 178 kin-pairs (HSPs and POPs) rather than on 76 (POPs alone). Although internal CVs are lower than in the original Bravington et al, 2014 model – i.e., precision is better – the reduction is not as great as one might expect based purely on the increase in number-of-kin-pairs. The reasons for this are: (i) the new model is more flexible and more parameters are being estimated, and (ii) the LSfreq data is handled differently in the new model and its contribution will be less than in the original study, so it contributes less apparent precision. The range of point estimates from alternative model assumptions is now of the same order of magnitude as the reported CVs, and it appears (from the evidence of some internal tension between the datasets; see below) that the LSfreq is still being weighted too highly. Thus, the internal CVs must still

be somewhat low, and more exploration is warranted. This was less obvious in 2013, perhaps because fewer options could be explored, and the CVs were larger anyway because of reduced sample size.

Model fits and Diagnostics

- All reasonable model options give reasonable to good fits to a range of close-kin summary statistics (see section below). So far, the best fits to CKMR statistics have been obtained with fixed selectivity; free-selectivity models do show some tension between POPs and HSPs, presumably reflecting tensions between CKMR and LSfreq data (or the handling of the latter in the model).
- All options fit equally badly to the sex-ratio in the Indonesian LSfreq data, in particular showing less trend (towards more males) than is seen in the observed data;
- The fits are compatible with $\alpha_{\text{HSP}} = 1$, which would in fact be the point estimate if the two sexes were constrained to have the same value of α_{HSP} . This is a “no surprises” result; the representation of individual fecundity appears consistent with the CKMR data, and the POPs and HSPs are telling the same story. Given the current totals of about 75 POPs and 140 HSPs, the lower confidence interval on α_{HSP} (which is set by the ratio of those totals) is about 0.85; thus, values slightly below 1 are not ruled out, but there is no evidence for them in the observations. In particular, there is no evidence for “hidden population structure”, whereby some adults persistently tend to have offspring that go to South Africa rather than the Great Australian Bight (GAB). CKMR alone can never exclude the possibility that some juveniles do go to South Africa, rather than to the GAB, but there is no evidence that their siblings are also more likely to do so.
- When selectivity is estimated and $\alpha_{\text{HSP}} = 1$, there is some mismatch between the POP and HSP totals – not statistically significant, but nevertheless noticeable. Letting α_{HSP} be estimated would not alleviate matters, since the preference of the model would be to make the estimate $\hat{\alpha}_{\text{HSP}} > 1$ which is not possible (see below). In other words, the best practical fits with estimated selectivity still have $\alpha_{\text{HSP}} = 1$.

One unreasonable model option is to simultaneously allow estimated α_{HSP} and estimated selectivity. The problem is that the fundamental information for either parameter is encapsulated in the same number--- the ratio of total-HSPs-to-total-POPs--- so you can estimate one or the other but not both. Basically: the POPs determine the total amount of reproductive output, but do not indicate whether that comes from fewer older (bigger) adults or from more younger (smaller) ones. However, fewer older adults means that “family sizes” are larger, and there will be more HSPs overall (since the number of HSPs in a “family” is quadratic in the size of the family)--- so the ratio of HSPs to POPs is largely set by the average age of living adults. The latter also determines the main “slope” of the Indonesian selectivity curve, which has to link the observed length frequencies of adults to the numbers-at-age in the population. Alternatively, if selectivity is treated as known (i.e. making the same assumption about residence-time that was used in the 2012 POP-only model), then the ratio of HSPs-to-POPs becomes available for other estimation purposes, e.g. for checking α_{HSP} . In fact, it is not just the totals that are relevant--- there are time-series aspects to numbers-of-POPs and numbers-of-HSPs--- but the main statistical information comes from the ratio of totals, and so it is not reasonable to estimate two quantities from the same number⁹.

⁹ did actually attempt exactly that (before realizing why it did not make sense) and the result was totally unreasonable estimates of α_{HSP} (around 2 or 3, which is completely opposite to any GAB-specific biological hypothesis)--- which, with hindsight, is not at all surprising.

Similar results were obtained from the POP+HSP OM in Hillary et al (2017c) (bearing in mind that the OM is actually formulated in terms of a different parameter, $q_{hsp} = 1/;$ the OM results are here reinterpreted in terms of α_{HSP}). That OM also preferred α_{HSP} somewhat above 1, but again the results were consistent with the maximum plausible, and most desirable, the case of $\alpha_{HSP} = 1$. Since there is no strong *a priori* reason to expect $\alpha_{HSP} < 1$ (i.e. parental “preference” for sending offspring East or West several years after birth) and also no actual evidence that it is below 1, despite reasonable sample sizes (numbers of POPs and HSPs), it seems fair to assume a "base case" of $\alpha_{HSP} = 1$ at least until there is any clear evidence to the contrary.

Fits to A@LS data are good across options (Figure 7). Fits to the length-frequencies LSfreq look very impressive for all options (not shown). Unfortunately, the plots actually contain no useful diagnostic information, because the annual selectivity adjustments adjust automatically to match the observed data quite closely. In fact, there must be substantial differences in how hard the annual adjustments are having to work, leading to statistically better fits for certain options, but these are not easy to see; some better way of plotting the diagnostics is needed here. Fits to sex frequency in LSfreq, however, are strikingly bad for all options despite the annual selectivity adjustments, which so far are not sex-based (Figure 8).

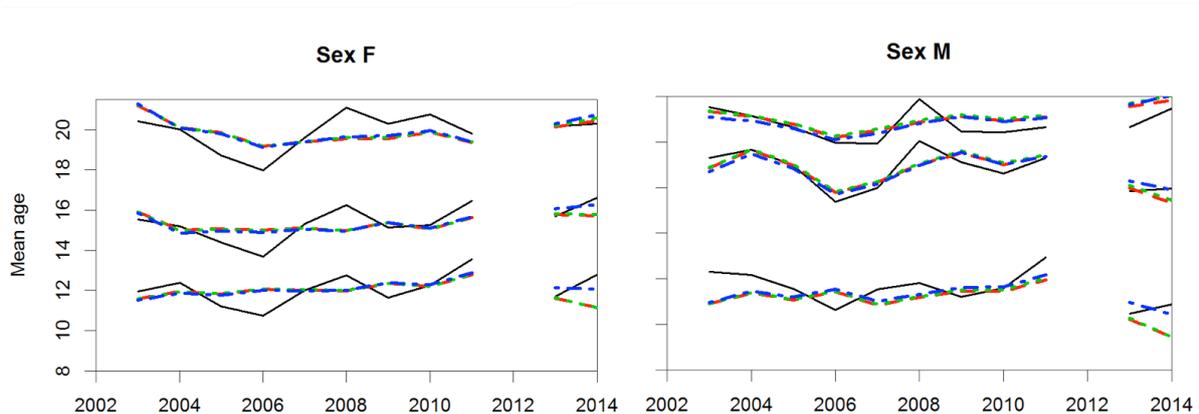


Figure 7: Fits to A@LS data, for females and males. Thick black lines are observed mean ages; coloured and broken lines are predictions from different options. The three groups of lines are: top, for big fish i.e. with lengths in the upper tercile (third) of all sampled lengths; middle, all fish; bottom, for small fish whose lengths are in the lower tercile. 2002 is missing because of concerns about the reliability of sex measurements in 2000–02. 2012 is missing because age estimates from otolith-reading are not available for that year.

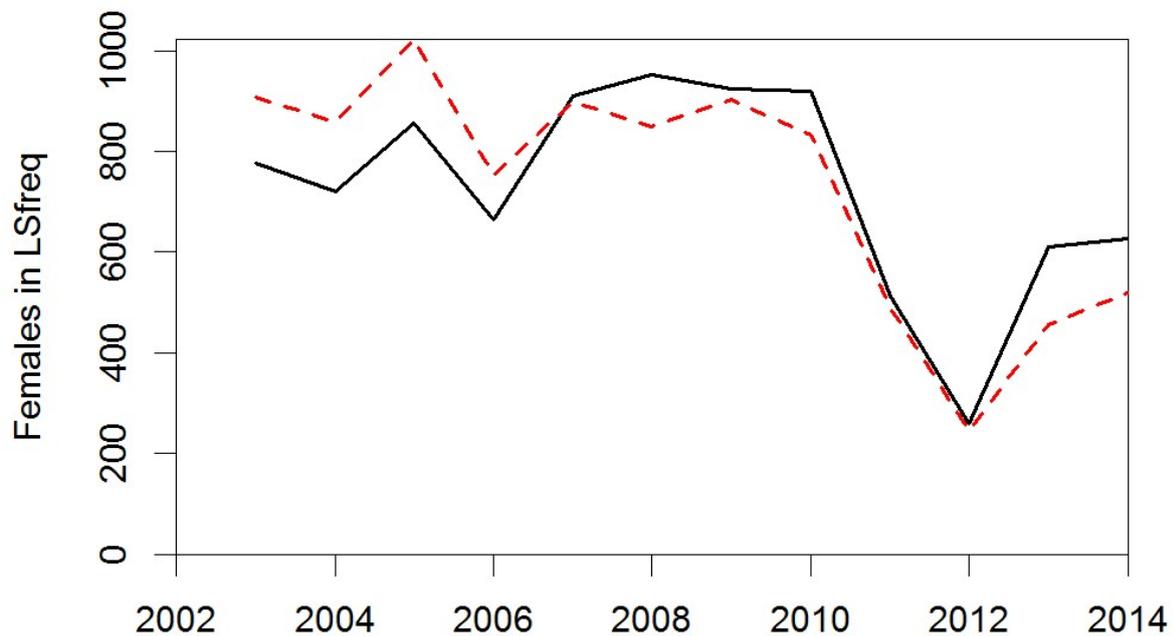


Figure 8: Fits to total number of females in LSfreq data. The thick black line is the observed count; broken pink line is the prediction (allowing for total sample size that year). All model options give essentially the same prediction, so only one is shown. Absolute numbers have been plotted to show the very considerable magnitude of the discrepancy (6 Standard Deviations in many years).

CKMR diagnostics

The match between the observed and predicted CKMR summary statistics seems generally good. Table 5 shows some simple summaries. Predictions within ± 1 SE of the observation are entirely consistent with the data, and between about 1 and 2SE are just about consistent; above two SE indicates misfit. Fits to males are worse than to females (and less effort has been spent trying to represent male fecundity in the model). In all options, male turnover rates (corresponding to Pdt in the Table) are predicted somewhat too low, and the male parental length is predicted significantly too high. This latter difference is mitigated when the LSfreq data is downweighted. Other than that, the only noteworthy feature of these summaries is the moderate mismatch between HSP and POP totals in option Bc (free selectivity, full weight to LSfreq).

The one badly-fitting option

The one really bad fit to CKMR data occurs when selectivity and α HSP are both estimated. In theory, this should give the best fits because it has the most flexibility to adjust parameters, but in practice it gives the worst! The problem is that the point estimate of α HSP rises to about 2, for both sexes; this about 3 standard deviations above the maximum plausible value of 1, so clearly is not just an unlucky accident of data. This option leads to only slight improvements in the fit to the observed CKMR statistics (α HSP being a parameter not an observed statistic), so presumably the real gain is somehow in an improved fit to LSfreq.

Unsurprisingly, such a high α does markedly change the abundance estimates, but since the parameter estimates make no sense, this result can be disregarded. Nevertheless, it does indicate some unresolved tension between datasets.

It seems likely that selectivity in the LSfreq data is the root cause of misfit; some more work is needed here. The LSfreq data has very large sample sizes, and unless carefully handled it tends to bully the other datasets to get its own way in the log-likelihood; that is, the model ends up willing to sacrifice appreciable goodness-of-fit to CKMR and A@LS data in order to accommodate minor nuances of the LSfreq data (although the models checked so far are still unable to match the sex proportions in the LSfreq, as mentioned). In principle, this dominance should be eliminated by the annual selectivity adjustments, but evidently our attempts at that have not yet met with complete success. There are two likely reasons:

- the basic model for selectivity and how it varies might not yet be formulated appropriately, especially with respect to sex;
- the variance of the year-to-year fluctuations in selectivity has been pre-estimated (see Appendix) and fed into the entire model as a fixed parameter. This would be OK if the pre-estimated variance was appropriate, but in fact it seems that rather more variance is required to match the data, especially with respect to sex frequencies. In particular, the pre-estimated variance was based only on year-to-year changes in length-frequency, not sex-frequency. The statistical software used to fit the model currently has the limitation that that only one random-effect variance parameter can be estimated inside the model; any other random-effect variances must be supplied a priori as fixed values. Currently, that one variance has been reserved for recruitment variability, so the annual selectivity variance has to be pre-estimated. It is preferable, at least in theory, to estimate all random-effect variances inside the model, and future versions of the software will eventually allow this.

Table 5: CKMR summary statistics: observed and expected, by sex, for each combination of selectivity (A/B: fixed/free selectivity) and length-frequency data-weighting (c/d: full weight/down weight). All options in this table have α HSP fixed at 1. Hdt: interval between birth-year in cross-cohort HSPs. Htot: the total number of HSPs. Pdt: interval between offspring birth and parental recapture. Pl: length of parents at capture. Ptot: the total number of POPs dSE: number of standard errors (computed empirically from observations) between prediction and observation.

	Hdt				Htot			Pdt				Pl			Ptot					
	F	dSE	M	dSE	F	dSE	M	dSE	F	dSE	M	dSE	F	dSE	M	dSE	F	dSE	M	dSE
Obs	3.07		3.05		45		57		3.56		3.33		171		175		36		40	
Ac	2.92	-0.5	2.96	-0.4	47.7	0.4	57.4	0.1	3.65	0.2	3.73	1.4	173	1.4	179	4.4	33.4	-0.4	39.7	0
Bc	3.04	-0.1	3.05	0	53.4	1.3	65.3	1.1	3.58	0.1	3.68	1.2	173	1.7	180	5.1	27.6	-1.4	31.7	-1.3
Bd	3.23	0.5	3.22	0.7	50.5	0.8	58.4	0.2	3.88	0.8	3.93	2.1	171	0	176	1.4	30.5	-0.9	38.6	-0.2
Ad	3.14	0.2	3.11	0.2	47.8	0.4	55.9	-0.1	3.84	0.7	3.88	1.9	171	0.2	177	1.8	33.2	-0.5	41.1	0.2

Conclusion

The overall goal of this project was to improve the accuracy of the stock assessment for Southern Bluefin Tuna by providing a fisheries independent estimate of absolute spawning biomass from a second application of the close-kin mark-recapture approach and POP and HSP data for inclusion in the CCSBT Operating Models used for period assessments of the state of the stock. This was achieved through this, and two related (CSIRO and CCSBT) projects, by successfully:

- processing and genotyping ~16,000 archived tissue samples to identify 77 Parent-Offspring and ~140 Half-sibling pairs;
- providing these data (POPs, HSP and related comparisons) for inclusion in the CCSBT operating models for the 2017 international stock assessment;
- estimating a decadal times series of absolute spawning potential of the SBT population using the new POP and HSP data, the extended CKMR modelling framework and the age and length composition data of the Indonesian catches from the spawning ground;
- including the CKMR data and parameter (false-negative rate) in CCSBT OMs, which reduced the overall structural uncertainty across the grid of operating models, including natural mortality for adults¹⁰, and;
- presenting the results to industry, AFMA, ABARES, Fisheries Section of DAWR and CCSBT for review as they became available to ensure their uptake at the national and international level.

Overall, the delivery of the combined outputs of this and the related projects has resulted in the new CKMR data being an agreed input in to the 2017 international stock assessment and an agreed monitoring series for the development of candidate Management Procedures for CCSBT; the CCSBT has committed to ongoing funding for the collection of adult (Benoa, Indonesia) and juvenile (Pt Lincoln, Australia), processing and kin identification as part of the CCSBT Scientific Research Program; and the Scientific Committee has recommended that the stand-alone CKMR assessment be repeated on a periodic basis for comparison with and testing assumptions of the CCSBT Operating Models.

The following general points are worth noting in the context of the successful delivery of this project:

Cost-effectiveness of the move to SNPs

The investment by CSIRO (see Appendix 3) in exploring alternative Next-Generation sequencing platforms and development of specific sequencing approaches for SBT (specifically, DArTcap) has returned significant benefits, both in terms of cost, speed and repeatability/quality control. In comparison to the original SBT application using specifically designed micro-satellites, the cost of marker “discovery/development” is ~25% for SNiPs and the cost of large-scale sequencing is ~50% or less. This makes the use of CKMR a cost-effective proposition for a wide range of fisheries and conservation management species (e.g. Feutry et al, 2017; Hillary et al 2018). In the case of SBT, the CCSBT is now collecting, sequencing and completing kin-identification to provide the data for regular updates of CCSBT Operating Models, use in Management Procedures and CKMR stand-alone assessments for less than \$100k/annum.

¹⁰ Anon (2017). Report of the OMMP Technical Webinar on the incorporation of half-sibling pairs in the CCSBT OMs for the 2017 update of stock status. Working paper CCSBT-ESC/0817/36 to the Twenty-Second Meeting of the Extended Scientific Committee. Yogyakarta, Indonesia, 28 August – 2 September 2017.

https://www.ccsbt.org/en/system/files/ESC22_36_report_2017HSPWebMeeting_0.pdf

Anon (2017b). Report of the Twenty-Second Meeting of the Extended Scientific Committee. Yogyakarta, Indonesia, 28 August – 2 September 2017.

https://www.ccsbt.org/sites/default/files/userfiles/file/docs_english/meetings/meeting_reports/ccsbt_24/report_of_SC22.pdf.

Increased information content from a combination of POPs and HSPs

The primary benefit of the move to SNPs is the power to resolve more distant kin, including full siblings and half-sibling pairs, and the analysis of mitochondrial DNA to identify the sex of parents of kin pairs. The addition of half-sibling pairs, in combination with the POPs, not only provides the basis to separately estimate total adult mortality and selectivity, it allows a range of potentially important assumptions to be tested. For example, whether the rate of turn-over of the total reproductive potential of the population varies by sex, or whether there may be a spatial or temporal structure in the distribution of juveniles. For example; do some subset of adult SBT produce juveniles that systematically go to the Great Australian Bight, while another produce offspring that go to South Africa? While, in the SBT case, the results do not indicate the presence of such “hidden” population structure, the ability to test this form of assumption directly is of considerable general benefit, as: i) it is very difficult to detect and directly measure this sort of population structure by other means, and ii) it has significant implications for the effectiveness of monitoring and management systems.

Implications

The outputs from this and the related projects, which made its delivery possible, have SBT-specific and more general implications for monitoring and management of fisheries. In the case of SBT, it has demonstrated the power and utility of the POP+HSP method of CKMR to estimate three of the central parameters for fisheries stock assessment: an abundance of adults, total mortality and selectivity of the adult component of the SBT stock. This has been done in a stand-alone CKMR assessment framework that is independent of the catch and effort data for the major longline fisheries, and the large uncertainties associated with the interpretation of the CPUE series derived from them. These data and the stand-alone assessment have substantially reduced the uncertainty in the state of the stock, improved the stability of the CCSBT OMs used for stock assessment and, as a direct result, substantially increased national and international confidence in the estimates of stock status. In addition, the time-series of POP and HSPs have been shown to be highly informative when used in Candidate Management Procedures (CMPs) designed to set the level of global catches of SBT.

As a result, the Australian Government, stakeholders and the CCSBT have agreed to the use of these two data series (POPs and/or HSP) in CMPs being developed and tested in the CCSBT Management Strategy Evaluation process. A final Management Procedure (MP) will be selected by the CCSBT to replace the current MP and used to set the TAC for 2020 and beyond. Importantly, this science has been developed and delivered in a manner that has engaged both national stakeholders and managers and members of the international management organisation, which has resulted in substantive “buy-in” and ongoing support. This has direct ongoing benefits for the Australian Government, industry and wider stakeholders, as well as for the other members of the CCSBT.

In the wider fisheries stock assessment and management context, this project has delivered the first application of what is likely to become the “template” method of CKMR for most teleost populations. The strong non-linear increase in fecundity with size/age in most teleosts means it will be necessary to use the POP+HSP method to generate unbiased and reasonably precise estimates of adult abundance and test the underlying assumptions of CKMR. The shift from microsatellite (used in the original study) to SNP markers for this project, combined with assay developments and streamlining of laboratory work-flows by CSIRO, has demonstrated it is possible to efficiently process large sample sizes (17,000 individuals in ~ 6 months) and identify HSP sufficiently accurately to provide the required samples sizes for precise estimates of abundance and mortality. These empirical results are consistent with those expected from CKMR theory and, combined with positive results from CKMR studies for other species, continues to demonstrate the breadth of utility and cost-effectiveness of the approach for fisheries and conservation management.

Recommendations

Recommendation from this project to inform decision makers:

The original application of Close-kin Mark Recapture had made substantial contribution to reducing the uncertainty in the status of the SBT stock and other aspects of the operating models used by CCSBT. This second project, using SNP marker and POPs and HSPs in combination, has further reduced the uncertainty in stock status and, importantly, demonstrated that large-scale sample collection and genotyping can be routinely applied to a large-scale, internationally managed fishery, such as SBT.

In light of this we recommend:

1. The ongoing collection of samples (tissue, otoliths and lengths) from Benoa (Adults) and Port Lincoln to provide the basis for i) regular use in the CCSBT Operating models for assessments of stock status (currently every 3 years); ii) periodic, updates of the CKMR (~5years), and iii) inputs to future MPs.
2. Given the collective value of the CKMR data to the CCSBT monitoring, assessment and management, the collection, processing and genotyping for CKMR be funded by the CCSBT through its Scientific Research Program.
3. That the CCSBT Scientific Committee consider the incorporation of CKMR data (HSPs and/or POPs) as a core data series for use in the development of Candidate Management Procedures for recommending the global TAC.

Recommendations for further research:

The impact of this work for SBT and other species would be enhanced through:

1. Continued refinement of lab workflows to reduce cost, time and increase quality control of the resulting genotypes and kin relationships. This would have wider benefit to a range of future applications beyond SBT.
2. Refinement of software and genotyping methods to increase resolution for identifying HSP and more distant kin. This will improve accuracy and reduce longer-term cost.
3. Ensuring long-term viability and consistency of adult (Indonesia) and juvenile (Australian) sampling programs.

The value and impact of CKMR to assessment and management of Australian Fisheries and wildlife management more broadly would be increased by:

4. Completing a review of the applicability of CKMR to a range of life-history and fishery types and developing guidance for the design and implementation of CKMR studies for Australian fisheries. This would share expertise, assist in building understanding of the approach and focus funding and applications in priority areas where CKMR is likely to provide the most value.
5. Building quantitative capacity for the wider application of the CKMR approach to Australian and other fisheries. A first step towards this would be the development of workshop materials and design software for applied fisheries and conservation researchers. A second step would be further development and generalisation of current “bespoke” code for CKMR design, kin identification and abundance applications into open-source packages that are accessible to a wider range of practitioners.

Extension and Adoption

This project has been strongly supported by stakeholders and management at the national level since the successful completion of the original proof of concept application of CKMR to SBT (Bravington et al, 2014; 2016). The proposal for this project was developed directly with the support of ASBTIA as a recognised priority and supported by AFMA, SBTMAC and, in particular, ABARES and the International Fisheries Section of DAWR. The Principal Investigator for this project (Davies) and for related SBT inter-sessional projects (Preece) are observers to SBTMAC and this project was identified and agreed as a priority for the fishery at a national level in 2015.

At an international level, the Principal Investigator leads CSIROs engagement in CCSBT and worked closely with the Australian Heads of Delegation to the Scientific Committee to re-initiate the CCSBT Scientific Research Program (Stobutzki et al, 2013) and engage other members of CCSBT and the Advisory Panel in the review of priorities and re-commitment of funding to strategic research needs for the Commission. Strategic design studies for the use of CKMR were identified as priorities and funded by CCSBT under this program (Bravington and Davies, 2013; Bravington, 2014, Bravington et al, 2015), as were the latter years of collection and archiving of tissue, which provided the necessary time series of samples for this project (Anon, 2013, 2015; Farley et al, 2017).

The outputs of the project have been formally adopted by the CCSBT and incorporated under the CCSBT Scientific Research Program as part of the ongoing monitoring of the spawning stock, as inputs to the regular stock assessment and to Candidate Management Procedures currently undergoing testing in the CCSBT. It is quite likely that the new MP selected for implementation by the CCSBT for setting the global TAC beyond 2020 will include close-kin data¹¹.

These SBT related outcomes have only been possible due to the support from all stakeholders at the national and international level for the long-term monitoring and sample collection in Indonesia and Australia, the initial design work that underpinned this project and the support and cooperation for the implementation of this project.

Table 6: Summary of consultation, extension and adoption at national and international level associated with the development and delivery of this FRDC project. References correspond to those in the main text.

Year	Event - outcome	Reference
2013	CCSBT-ESC: <ul style="list-style-type: none"> Review of original CKMR application to SBT complete and POP data incorporated into CCSBT Operating Models. Strategic paper on the future developments and potential value of CKMR for monitoring and assessment of SBT. Design study for CKMR included under CCSBT Scientific Research Program SBTMAC: <ul style="list-style-type: none"> Value of CKMR results to increasing confidence in stock status strongly acknowledged by Industry and Management CSIRO and Industry supports ongoing collection of samples. 	Bravington et al 2012, 2014 2016; Hillary et al 2012, 2013; Anon, 2013 Bravington and Davies 2013 Stobutzki et al 2013, Anon 2013
2014	Industry consultation – Pt Lincoln <ul style="list-style-type: none"> Final CKMR results from ESC 2013 and implications presented CCSBT- ESC: <ul style="list-style-type: none"> Evaluation of SNP versus micro-satellites markers and preliminary design calculations for POPs+HSP using SNPs ESC recommended independent advice on genetics of HSP method and selection of 	Bravington 2014 Anon, 2014

¹¹ At the 2019 meeting of the CCSBT Extended Scientific Committee recommended to the Commission a single Candidate MP, which included the CKMR, gene-tagging and longline CPUE monitoring series (rh12, Hillary et al 2019, Anon 2019). The Commission adopted this MP, now known as the “Cape Town Procedure, at its 26th Annual meeting in Cape Town, South Africa.

	<p>genotyping platform</p> <p>SBTMAC:</p> <ul style="list-style-type: none"> Review of research priorities identifies future application of CKMR as high priority 	
2015	<p>Industry Consultation – Pt Lincoln</p> <ul style="list-style-type: none"> Summary of sampling and re-affirm priority of CKMR as a fisheries independent estimate of stock status and input to 2017 stock assessment <p>CCSBT- ESC:</p> <ul style="list-style-type: none"> Expert review of proposed change in sequencing platform Updated design study for POP+HSP method using SNPs Review of priorities for CCSBT-SRP New CKMR study a priority for 2017 stock assessment along with a pilot study for gene-tagging to provide recruitment index CCSBT agreed to fund processing of 2 years of samples <p>SBTMAC:</p> <ul style="list-style-type: none"> SBTMAC recommend CKMR study as high priority for FRDC funding 	<p>Anderson and Waples 2015</p> <p>Bravington et al 2015</p> <p>Anon 2015a</p> <p>Anon 2015a</p> <p>Anon 2015b</p>
2016	<p>Industry Consultation – Port Lincoln</p> <ul style="list-style-type: none"> Overview of CKMR POP+HSP approach, project schedule and Industry support for sampling CSIRO commitment to CKMR model development to use data from CCSBT and proposed FRDC project EoI submitted to FRDC, Project approved Aug 2016, Project activities commence Oct 2016 <p>CCSBT- ESC:</p> <ul style="list-style-type: none"> Report on progress with CCSBT CKMR funded project Verbal report on FRDC project status 	<p>Davies et al 2016</p> <p>Farley et al 2016</p>
2017	<p>CCSBT- ESC:</p> <ul style="list-style-type: none"> New POP and HSP data provided to and reviewed at OMMP Technical meeting, June 2017, Seattle. POP data and new HSP data incorporated into OMs and reviewed as special Webinar of ESC in August, 2017. POP and HSP data provided and included in updated stock assessment and advice on stock status in September 2017 Approach to CKMR stand-alone model reviewed by ESC ESC agrees to include CKMR data as core monitoring series for development of new Candidate Management Procedures ESC recommends (and Commission agrees) to funding ongoing collection, processing and kin identification for CKMR. <p>SBTMAC:</p> <ul style="list-style-type: none"> June, Summary of progress to date including data exchange with CCSBT for OMMP Technical meeting. September, Briefing on project status and outcomes of inclusion of new POP and HSP data in OMs. <p>Industry Consultation – Port Lincoln Research Day, November</p> <ul style="list-style-type: none"> Full over-view of ESC meeting including CKMR project and results on inclusion of CKMR data in OMs for assessment of stock status. 	<p>Bravington et al 2017a</p> <p>Hillary et al 2017a, Anon 2017a</p> <p>Bravington et al 2017b, Hillary et al 2017b, Anon 2017b</p> <p>Anon 2017b</p> <p>Anon 2017b</p>
2018	<p>CCSBT- ESC:</p> <ul style="list-style-type: none"> Full technical report, including stand-alone CKMR assessment submitted to ESC. POP and HSP data formally exchanged as part of the CCSBT-ESC data exchange Ongoing funding of CKMR and Gene-tagging data streams under CCSBT Scientific Research Program. <p>Recognition:</p> <ul style="list-style-type: none"> CSIRO SBT Team (and collaborators) awarded: i) CSIRO Medal for Impact through Science and ii) Sir Ian McLennan Award for Achievement for Industry for SBT stock assessment, MP, CKMR and Gene-tagging. Industry, Conservation, Management, Policy, FRDC and CCSBT attend Award ceremony. 	<p>Davies et al 2018</p> <p>Anon 2018</p>

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Project materials

The following working papers from this project were submitted to the CCSBT:

1. Bravington, M.V. (2017). SBT kin-finding and genotyping update. Paper to the 8th meeting of the Operating Model and Management Procedure Technical Group of the CCSBT, 19-23 June 2017, Seattle, U.S.A. (CCSBT-OMMP/1706/12). 11p.
https://www.ccsbt.org/en/system/files/OMMP8_12_AU_CKMR_summary-ommp-seattle-2017.pdf
2. Bravington, M.V., Eveson, J.P., Grewe, P.J and Davies, C.R. (2017). SBT Close-Kin Mark-Recapture with Parent-Offspring and Half-Sibling Pairs: update on genotyping, kin-finding and model development. Paper CCSBT-ESC/1706/12 prepared for the Extended Scientific Committee for the Twenty-Second Meeting of the Scientific Committee, Yogyakarta, Indonesia, 28 August – 2 September 2017. 29p.
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3. Davies, C.R., Bravington, M.V., Grewe, P.J. and Eveson, J.P. (2018). Close-kin project report. Paper CCSBT-ESC/1809/14 prepared for the Extended Scientific Committee for the Twenty Third Meeting of the Scientific Committee, San Sebastian, Spain, 3-8 September 2018.

Related Project materials

The following working papers from the related projects were submitted to the CCSBT:

1. Hillary R.M., Preece, A. and Davies C.R. (2017a). Updates required for new data sources and reconditioning of the CCSBT OM. Paper (CCSBT-OMMP/1706/4) to the 8th meeting of the Operating Model and Management Procedure Technical Group, 19-23 June 2017, Seattle, U.S.A. 19p.
2. Hillary R.M., Preece, A. and Davies C.R. (2017b). Summary of initial incorporation of the Half-Sibling Pair (HSP) data in the CCSBT Operating Model. Paper CCSBT-OMMP/1706/03 to the CCSBT-OMMP webinar, 21 July 2017. 8p.
3. Hillary, R.M., Preece, A.L., Davies, C.R., Takahashi, N., Sakai, O. and Itoh, T. (2017). Reconditioning of the CCSBT Operating Model in 2017. Paper CCSBT-ESC/1706/14 prepared for the Extended Scientific Committee for the Twenty-Second Meeting of the Scientific Committee, Yogyakarta, Indonesia, 28 August – 2 September 2017. 25p.
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Committee for the Twenty Third Meeting of the Scientific Committee, San Sebastian, Spain, 3-8 September 2018. https://www.ccsbt.org/en/system/files/ESC23_08_CCSBT_CKMR.pdf

6. Hillary R.M., Preece, A. and Davies C.R. (2018a). Data generation & changes to SBT OM. Paper CCSBT-ESC/1809/19 prepared for the Extended Scientific Committee for the Twenty Third Meeting of the Scientific Committee, San Sebastian, Spain, 3-8 September 2018. https://www.ccsbt.org/en/system/files/ESC23_19_AU_ProjChangestoOM.pdf
7. Hillary R.M., Preece, A. and Davies C.R. (2018b). Performance of Revised CMPs. Paper CCSBT-ESC/1809/20 prepared for the Extended Scientific Committee for the Twenty Third Meeting of the Scientific Committee, San Sebastian, Spain, 3-8 September 2018. CCSBT-ESC/1809/20 https://www.ccsbt.org/en/system/files/ESC23_20_AU_MPpaper_ESC.pdf

Appendices

Appendix 1: Technical description of Close-kin Mark Recapture abundance estimation model for Southern Bluefin Tuna

Appendix 2a: CSIRO project staff

Appendix 2b: CSIRO staff - related projects

Appendix 2c: Supporting agencies and companies

Appendix 3: Related Projects

Appendix 1: model details, SBT CKMR 2018

Parameters are estimated within a REML framework, using Laplace Approximation and automatic differentiation to approximate the marginal log-likelihood allowing for random-effects. This is the same statistical framework used in e.g. TMB (Kristensen et al., 2016) and ADMB-RE (H. J. Skaug and Fournier, 2006), but implemented via our in-house ADT software, which is far easier to debug. There is an outer layer of 3000 lines of R (R Core Team, 2013) code to organize the data, set up the model options, oversee parameter estimation, and extract summary statistics. The inner code, which implements the population dynamics and computes the log-likelihood from the observed data, consists of about 1500 lines of Pascal code (similar to C), about half of which is “housekeeping”. Efficient derivative computation is crucial for Laplace Approximation, and this is achieved by automatic differentiation using the source-code transformation tool Tapenade (Hascoët and Pascual, 2013); our in-house ADT software organizes the calls to Tapenade. Runtime to fit a model is 5–10 minutes, and no numerical problems have been seen during fitting.

Particular model options, e.g. which covariates are allowed to affect which parameters, are implemented through R’s standard syntax for statistical formulae, so it is possible to explore many different options without touching the underlying Pascal code. For example, most of the options follow the current OM practice in assuming that survival probability¹ should be independent of sex and year, constant between ages 8 and 24, and then following a linear trend (actually on a logistic scale) to meet the plus-group z at age 30. This can be specified like so:

```
logit_psurv_formula = ~ I( pmax( age, 24)- A_MAX) - 1
```

To allow dependence on sex, the formula would become:

```
~I(pmax(age, 24) - A_MAX) %in% sex - 1
```

In the descriptions below, subscripts are used to indicate which covariates are *potentially* allowed to affect which variables. In most options, only a subset of the potential covariates are allowed, and this is noted.

¹In fact, the formula applies to the transformed mortality rate, $\text{logit}(e^{-z})$, and there is an implicit offset of $\text{logit}(\exp(-z_{A_{\max}}))$, the plus group survival, discussed below.

The rest of this Appendix presents details of the population dynamics, and treatments of the four datasets, following a table of notation. For clarity, the words “we assume that...” are mostly omitted, but should be read implicitly throughout; e.g., the text *states* that growth follows individual-specific von Bertalanffy curves, but what is meant is that we *assume* that to be the case and have coded the model accordingly.

Population dynamics

Population dynamics follows a standard sex- and age-structure with annual time steps, using mortality rates but no explicit catches:

$$\begin{aligned} n_{s,y+1,a+1} &= n_{sya} \exp(-z_{sya}); \quad a < A_{\max} \\ n_{s,y+1,A_{\max}} &= (n_{s,y,A_{\max}} + n_{s,y,A_{\max}-1}) \exp(-z_{s,y,A_{\max}-1}) \end{aligned}$$

which automatically imposes the constraint that plus-group survival is the same as in the preceding age-class. No year-dependent options have actually been tried yet. Plus-group age A_{\max} was set to 30, except when repeating the 2013 analysis where 25 was used; data-wise, all measured ages above A_{\max} were treated as “30 or more”.

For CKMR probabilities, it is necessary to keep track of *some* information about ages within the plus-group. The mean age within the plus-group evolves according to :

$$\bar{a}_{\max,s,y+1} = \frac{(\bar{a}_{\max,sy} + 1) \times n_{syA_{\max}} + A_{\max} \times n_{sy,A_{\max}-1}}{n_{sy,A_{\max}} + n_{sy,A_{\max}-1}}$$

and we assume that age-within-plus-group follows approximately an exponential distribution with that mean.

The 2013 analysis, without the benefit of HSPs, *required* the assumption of equal male and female survival rates by age. With HSPs, it is at least possible to relax that assumption. In limited testing so far, though, letting survival rate depend on sex has not made much overall difference, so all the options reported in the main paper have assumed equal male and female rates.

Minimum age is 8— i.e., animals enter the model only when they reach age 8, the first age with evidence of successful breeding.

“Recruitment” at age 8 is log-normally distributed around a constant mean. The ratio of of males-to-females at age 8 in the population is an estimated parameter, but it is assumed constant over time.

The CV of n_8 was estimated at 0.35 using 2002–2010 data (as in 2013 model). With 2011–2014 data

Table 1: Main notation. Many of these quantities are always disaggregated in practice by subscripts, e.g. to denote age.

Symbol	Meaning	Notes
y	Year; year-of-capture	Adults only
s	Sex	Adults only
ℓ	Length; length-at-capture	Adults only
b	Year of birth	Juveniles only
n	Numbers in population	
ϕ	Annual fecundity	Relative to the fecundity of a 170cm fish of that sex
σ	Selectivity	Chance of occurring in the LSfreq data, relative to a 170cm fish of that sex
z	Mortality rate	Annual survival probability= $\exp(-z)$
A_{\max}	Plus-group age	No growth and no age-specific changes in survival after that age
$\bar{a}_{\max sy}$	Mean age within plus-group	
m_P, m_H	Number of pairwise comparisons	m_P relates to POPs; m_H relates to HSPs.
i, j	Labels for individual fish involved in a pairwise comparison	
K_{ij}	Measured kinship of fish i and j	If i is adult and j is juvenile, then the possible values are POP or UP. If i and j are both juvenile, the possible values are definite MHSP, definite PHSP, or UP/false-negative-HSP.
#HSP	Number of HSPs found	
$\mathbb{E}, \mathbb{V}, \mathbb{C}, \mathbb{I}$	Expectation (mean); variance; covariance; indicator function	
\triangleq	“is defined as”	For temporary variables unworthy of inclusion in this table

1. The notation $\mathbb{P}[X | \{y\} z]$, for conditional probability of some event X given covariates y and z , means that covariate y is formally required in the conditioning because of the previous application of a probability manipulation such as Bayes’ theorem, but in practice is irrelevant to this particular probability.
2. “Prime” variables, such as a' for age, are used in summations to distinguish between separate occurrences of the same *type* of variable in a single formula.

added— a period which include some very strong 8yo recruitments— the estimated CV rises to 0.41. Different model options do seem to have much effect on the estimated CV of 8yo recruitment.

Plus-group survival rates

Catch-curve analysis of all post-1995 otoliths above age 30— an age above which growth has slowed enough that length-based selectivity should not affect the sampled age composition— shows an appreciably higher slope (CCSBT, 2009). These data were fitted in a preliminary analysis by a Poisson GLM with parameters β and z such that

$$\log \mathbb{E} [N_{sa}^{\text{oto}}] = \beta_{0s}^{\text{oto}} - z_{s,A_{\max}} \times (a - 30); a \geq 30$$

There is a suggestion from this analysis, albeit not statistically significant, that plus-group survival rates are lower in males than females.

Except when repeating the 2013 analysis, the estimate $\hat{z}_{s,A_{\max}}$ and its variance (by sex, if appropriate) from the preliminary analysis were incorporated as offsets in the main CKMR model, with the difference $z_{s,A_{\max}} - \hat{z}_{s,A_{\max}}$ treated as a random effect of known variance. In the 2013 model (and its reincarnation here), survival rate from age 25 up was estimated as a single parameter, and the composition of ages within the 25-and-up category was not used at all.

Initial age composition

In the first year of the model ($y_1 = 2002$), numbers-at-age are by default assumed to follow on average the catch-curve with slope ... (corresponding to constant average 8yo-recruitment in the past). These expected numbers in each age-class are then modified by cohort-specific random effects, just like incoming 8yo-recruitments in subsequent years. This is the same formulation as in 2013.

There is an option to estimate extra parameters for initial numbers-at-age, to allow general dependencies of the form $\log n_{s,y_1,a} | s, a$. This would accommodate, for example, changing pre-adult exploitation rates in the years before 2001, or (somehow) different sex frequencies in the initial spawning stock in 2002. Such options have not yet been explored, but might help deal with the mismatch to observed sex frequencies noted in the main text; nevertheless, it would be nice to have some insight into why such differences might have arisen.

Growth, length, and age

We have kept the 2013 formulation. Growth (only modelled for ages 8+) follows von Bertalanffy curves, with constant sex-specific k and t_0 for all adults of that sex, but individual-specific L_∞ that follows a Student's t_{12} distribution centred around a sex-specific mean (which is less sensitive than a

Normal to outliers from measurement error). Growth is deterministic, given an individual L_∞ . Formally, this is:

$$\mu_{sa} \triangleq \bar{L}_{\infty sa} (1 - \exp(-k_s(a - t_{0s})))$$

$$\mathbb{P}[\ell|sa] = F_{t_{12}}((\ell - \mu_{sa}) / (\text{cvl}_s \mu_{sa})) - F_{t_{12}}((\ell - \mu_{sa}) / (\text{cvl}_s \mu_{sa}))$$

where cvl_s is the CV of length-at-age and $F_{t_{12}}$ is the CDF of a standard Student's t_{12} distribution.

Since it is assumed that mortality rate depends on age rather than length, the population distribution (as opposed to sampling distribution) of length-at-age-and-sex keeps the same t_{12} -shape through the lifespan of each cohort, and individual fish maintain their “quantile” throughout adult life; in other words, a male SBT who was at the 15%ile of length-at-age-8 will still be at the 15%ile of length-at-age-28 twenty years later, if he survives. This considerably simplifies the calculation of HSP probabilities, described later.

Length classes are in 1cm intervals, with one grouped class at 150cm or below, and one grouped class at 200cm or above. For some calculations it is necessary to estimate contributions from animals below 150cm and above 200cm, so we also keep track of “mean conditional length” in those classes, e.g.

$$\mathbb{E}[L|L \leq 150\text{cm}, s, a]$$

which is computed analytically from properties of Student's t -distribution, as per formulae in BGD2016.

Age is assumed to be measured accurately, both for juveniles (all assumed to be age 3, based on all samples coming from a specific range of lengths) and adults. Otolith age estimates are known to be in error sometimes (usually by no more than 1 year), and it is also possible that some juvenile ages are wrong. In principle, ageing error can be accommodated in CKMR (and we have done so for other species), but it does greatly complicate the code. The effect of adult ageing errors on CKMR should not be that large; juvenile ageing errors would cause more problems, especially with mortality rate information from HSPs. The GT program should generate a lot more juvenile age-at-length data from tail vertebrae (a reliable indicator of age for immature SBT) so we will review that data when it becomes available.

Cohort-specific variations in growth have not been considered, though could be in principle.

Fecundity and maturity

We have kept the 2013 formulation; note that with the addition of HSPs, though, fecundity is no longer strictly coupled to selectivity (see below).

Males and females both reach maturity— i.e. to *potentially* breed successfully— at age 8. This is the youngest breeding age among the 76 POPs². For animals 8yo and up, relative fecundity depends on length (in a sex-specific way) but not on age per se. Female fecundity (annual) is proportional to spawning-ground residence-time multiplied by daily fecundity; the latter (in terms of egg biomass released) has been estimated, as a function of length, from the histological analyses. Residence time (on some relative scale) is then estimated within the CKMR model, assuming a logistic relationship to length.

Male fecundity is again proportional to spawning-ground residence-time multiplied by daily fecundity, but we have no prior information on the latter. CKMR only gives information on annual total fecundity as a function of body size (by sex), so it is not possible statistically to separately estimate male residence time and male daily fecundity. However, we continue to use residence time as the *basis* for selectivity (perhaps modified by other factors), so retaining the distinction is somewhat useful. As for females, male residence time varies logistically with length. Male daily fecundity has been assumed independent of length in most options, since it appears that any such relationship is imprecisely estimated; presumably, the logistic residence-time/length relationship may already provide enough flexibility to describe male annual fecundity, especially since selectivity can now be adjusted separately. (The overall male fecundity/size relationship was also difficult to estimate in 2013, but the direct linkage to selectivity complicated

The 2013 analysis suggested that skip-spawning (in alternate years) might be the norm for younger SBT adults but not for older ones, and the newer data broadly supports that. The average time-interval between offspring birth and parental capture has increased— simply because the data series spans more years— so the direct interpretation of birth/capture gaps is more fraught; an adult that was “young” when it bred may no longer be “young” when recaptured. Analysing the phenomenon properly in this larger longer dataset would require a bespoke statistical model (albeit much simpler than, and completely separate to, the main CKMR analysis); this would be an interesting exercise.

Skip-spawning is an aspect of fecundity which is not specifically allowed for in the CKMR probabilities described below. In a reasonably long study, such as we certainly have now, the effects should cancel out; about half the probabilities are calculated too low and about half are calculated too high, so for

²Actually, one parent in one post-2013 POPs does have an estimated age-at-breeding of 7. This is the only instance of an ostensible 7yo breeder in an enormous number of comparisons of 7yo adults (since each adult is involved in comparisons at all its ages between 2002 and the year before it was caught), while there are about six 8yo parents and far more at older ages; so, we suspect the 7yo estimate actually comes from otolith ageing error (estimated age 14 in this case). Even if this one instance is correct, it is abundantly clear that 7yo cannot make much of an overall contribution to SBT reproduction, so keeping age-at-maturity to 8 seems reasonable. Pushing down the age-at-maturity in the model introduces complications with length-frequencies that are just not worth the effort.

individual fish the effects should broadly cancel. At an aggregate level, the bulk of the reproductive output of SBT comes in any case from fish which are old enough to spawn every year.

While the CKMR framework of Bravington, Hans J. Skaug, and Anderson (2016) could certainly accommodate skip-spawning in principle, implementation would require much more complicated code. For SBT we expect there would be only minimal bias from using the current, much simpler, model.

POP equations

We have kept the 2013 formulation, which is a special case of the general ERRO (“Expected Relative Reproductive Output”) approach explained in BSA2016. In words: the chance of any given adult being the parent of a specific juvenile, is the expected reproductive output of that adult in the year and place that the juvenile was born, divided by the expected TRO (“Total Reproductive Output”) from all adults of the given sex at that year and place. The “expected” output needs to be calculated conditionally on all measured covariates for the adult; if the year and place of juvenile birth are not known but could take several values, then those values need to be integrated over conditional on what is known about the juvenile; and the art of CKMR is to express all that conditioning and integration correctly. For SBT, there is only one “place” (a single spawning ground) and juvenile age is (assumed) known, so all we need to worry about is the adult.

1. Fecundity $\phi(s, \ell)$ is driven by length not age (provided age is 8 or more), but since the length-at-age distribution is constant over time, we can also compute an age-specific average by

$$\phi_{sa} \triangleq \sum_{\ell} \phi_{s\ell} \mathbb{P}[\ell|sa]$$

where $\mathbb{P}[\ell|sa]$ comes from the distribution of growth curves across individuals.

2. Given the length, age, and sex of an individual adult in its year-of-capture³, its length $\ell^*(s, \ell, a, y, y')$ in any previous year-of-interest y' can be back-calculated straightforwardly. The equations are too dull to include, but the steps are:

- (a) work out its individual L_{∞} to match the observed length and age;
- (b) apply the individual growth curve at the age $a - (y - y')$ that the fish would have been in year y' .

For each combination of juvenile birth-year and certain adult covariates-at-capture (*not* including age, since age is only measured for a subset of genotyped adults), we compute probabilities based as

³There is not thought to be much length-growth during the spawning season, since adults lose weight on average, so date-of-capture within the season should not be important.

follows. The ERRO calculation takes into account whether the adult was likely to be alive and mature at the year of juvenile birth and, if so, its likely fecundity then:

$$\begin{aligned}\mathbb{P}[\text{POP}|bysl] &= \sum_{a=8}^{A_{\max}} \mathbb{P}[\text{POP}|bysla] \times \mathbb{P}[a|ysl\{b\}] \\ \mathbb{P}[\text{POP}|bysla] &= \frac{\phi(s, \ell^*(s, \ell, a, y, y-b)) \times \mathbb{I}[y > b+3] \times \mathbb{I}[a - (y-b) \geq 8]}{\text{TRO}_{s,y-b}} \\ \mathbb{P}[a|ysl] &= \frac{n_{sya} \mathbb{P}[\ell|sa\{y\}]}{\sum_{a'} n_{sy a'} \mathbb{P}[\ell|sa'\{y\}]} \\ \text{TRO}_{sy'} &= \sum_{a'} n_{sy' a'} \phi_{sa'}\end{aligned}$$

When turning this into a log-likelihood, the first step is to group the comparisons by alike covariates $bysl$, so that instead of nearly 100,000,000 individual comparisons, we need only compute about 70,000 probabilities. Only a tiny proportion of pairwise comparisons actually yield a POP. The computationally-efficient way to form the log-likelihood is to first compute it as if there were no POPs at all, then loop over the POPs individually to adjust the log-likelihood for the facts that (i) this comparison between adult i and juvenile j did yield a POP after all, and (ii) the age of adult i was measured to be whatever it is (since all the adults involved in POPs are deliberately aged). That is:

$$\begin{aligned}\Lambda_{\text{POP}} &= \sum_{bysl} m_{\text{P}bysl} \log(1 - \mathbb{P}[\text{POP}|bysl]) + \\ &\sum_{i,j: K_{ij}=\text{POP}} \text{logit} \mathbb{P}[\text{POP}|b_j y_i s_i \ell_i] \log(\mathbb{P}[a_i | b_j y_i s_i \ell_i, K_{ij} = \text{POP}]) \\ \mathbb{P}[a|bysl, K = \text{POP}] &= \frac{\mathbb{P}[\text{POP}|bysla] \times \mathbb{P}[a|\{b\}ysl]}{\sum_{a'} \mathbb{P}[\text{POP}|bysla'] \times \mathbb{P}[a'|\{b\}ysl]}\end{aligned}$$

HSP equations

This follows the principles explained in Bravington, Hans J. Skaug, and Anderson (2016). It is only necessary to compare *across* cohorts, not within them, so we can always distinguish the “first” and “second” juvenile in each comparison. Their joint HSP probability must be calculated by summing across all females (for MHSPs) or males (for PHSPs) alive at the time the first juvenile was born. Letting R_{ib} denote the *actual* (as opposed to expected or observed) reproductive output of animal i in year b , and supposing the two juveniles to be born in b_1 and b_2 respectively, the MHSP probability is

simply

$$\mathbb{P}[K_{12} = \text{MHSP}|b_1b_2] = \sum_{i \in \{\text{♀ alive at } b_1\}} \frac{R_{ib_1}}{\text{TRO}_{\text{♀}b_1}} \times \frac{R_{ib_2}}{\text{TRO}_{\text{♀}b_2}}$$

Note that the second output, R_{ib_2} , will be zero if the female i dies in-between b_1 and b_2 .

Given a large adult population, we can replace the actual R 's by their expected values for each female:

$$\mathbb{P}[K_{12} = \text{MHSP}|b_1b_2] = \sum_{i \in \{\text{♀ alive at } b_1\}} \frac{\mathbb{E}[R_{ib_1}R_{ib_2}]}{\text{TRO}_{\text{♀}b_1}} \times \frac{1}{\text{TRO}_{\text{♀}b_2}} \quad (0.1)$$

Clearly, it is the *covariance* between an individual's reproductive outputs which matters, not merely the product of the two expected values. The key to success with HSPs in CKMR is to subsume as much as possible of that covariance into specific covariates of adults (length, age, etc), so that the bulk of the “covariance” is encompassed into a sum of products of expected values, which themselves are computed as in the POP calculations. Any remaining unaccounted sources of covariance will evidently cause “bias” in the number of HSPs found. Almost all plausible mechanisms, e.g. infertility, would lead to positive covariance, so that HSPs will occur *more* frequently than a naive calculation would indicate, and a corresponding abundance estimate would be biased downwards.

Note that “purely random” variability in individual R in any single year, e.g. through “sweepstake reproduction” and “lucky litters”, does not matter; it is only *systematic* variability that needs to be accommodated. In the case of SBT, we can easily allow for adult length and growth by adapting the POP formulae; it would take a major mis-specification in our assumptions about growth trajectories (which are, basically, that bigger-than-average fish stay that way) to lead to much unaccounted covariance. Widespread infertility also seems a priori unlikely. Perhaps the most likely source of unaccounted covariance would be if some adults tend persistently to breed offspring who avoid the GAB in summer. To allow for such infelicities whatever their cause, the model can incorporate estimable scaling parameters α_{MHSP} and α_{PHSP} which act as multipliers on abundance when calculating HSP probabilities. If the model works as we hope, i.e. capturing the important sources of persistent individual-level fecundity, then these α_{HSP} should be close to 1. They might be substantially less than 1 if there are phenomena we have overlooked, so that there are more HSPs than “expected”. But they cannot reasonably be much greater than 1.

With SBT, the HSP formulae do need to account for systematic variation due to (i) adult age; (ii) individual growth curves within age, in that if You are at the 15%le of length-at-age in year b_1 then You will also be at the 15%le in year b_2 , and (iii) death. Point (ii) can be handled efficiently by summing across a fixed number Q of evenly-spaced quantiles of the t_{12} -distribution of length-at-age, so that $\{\ell(q\text{♀}a) : q \in \{1 \dots Q\}\}$ approximates an equiprobable set of lengths-at-age. The other terms are already available from the POP calculations. The overall probability for an MHSP becomes:

$$\begin{aligned}
\mathbb{P}[\text{MHSP} | b_1 b_2] &= \frac{1}{\alpha_{\text{MHSP}}} \times \\
&\sum_{a=8}^{A_{\text{max}}} \left\{ \frac{n_{\text{Q}b_1 a} \phi_{\text{Q}a}}{\text{TRO}_{\text{Q}b_1}} \times \right. \\
\frac{1}{Q} \sum_{q=1}^Q &\left\{ \frac{\phi(\ell(q_{\text{Q}a}))}{\phi_{\text{Q}a}} \times \phi(\ell(q_{\text{Q}}, a + (b_2 - b_1))) \right\} \times \\
&\exp \left(- \sum_{y=b_1}^{b_2-1} z_{\text{Q}y a + (y-b_1)} \right) \left. \right\} \times \\
&\frac{1}{\text{TRO}_{\text{Q}b_2}}
\end{aligned}$$

The extension to PHSPs is trivial, and we can also define $\mathbb{P}[\text{HSP}] \triangleq \mathbb{P}[\text{MHSP}] + \mathbb{P}[\text{PHSP}]$ since the two types of HSP are mutually exclusive; full-sibs across cohorts will be inconceivably rare in a random-mating population with millions of adults.

SBT have high haplotypic diversity in mtDNA, so much so that two HSPs which share an mtDNA haplotype are almost certainly an MHSP rather than a PHSP; if they have different haplotypes, of course, they must be a PHSP (see main text). This lets us treat the mtDNA evidence as definitive about MHSP vs PHSP, and simplifies the log-likelihood calculation quite a lot; for all other species that we have looked at, it has been both necessary and tedious to take into account the observed haplotypes for each HSP when computing the log-likelihood.

The final step for HSPs is to note that the measured kinship in HSP comparisons is not necessarily the true HSP status, but rather the fact of whether the PLOD (see main text) is above or below a certain threshold. That threshold is chosen on purely genetic grounds (i.e. before fitting the CKMR model) to eliminate any serious risk of false-positive ‘‘HSPs’’ from pairs that are unrelated or more weakly related. However—and in fact consequently—it is quite possible by chance that a true HSP will fall below the threshold and become a false-negative. This is accommodated by allow for a pre-estimated false-negative rate, in this case about 10%, so that the demographic HSP probabilities calculated above are all adjusted downwards to allow for false-negatives (i.e. reduced to about 90% of their nominal value) before computing the HSP log-likelihood, which is based on observed numbers of *definite* HSPs.

After all that, each pairwise comparison between juveniles is a ‘‘trinomial’’ event with outcomes MHSP/PHSP/UP, and the comparisons can be aggregated across birth-years to yield an overall HSP

log-likelihood as a sum of multinomials:

$$\begin{aligned} \Lambda_{\text{HSP}} = & \sum_{b_1} \sum_{b_2 > b_1} \{m_{\text{H}b_1b_2} \log(1 - \mathbb{P}[\text{HSP}|b_1b_2]) + \\ & \#\text{MHSP}_{b_1b_2} \log \mathbb{P}[\text{MHSP}|b_1b_2] + \\ & \#\text{PHSP}_{b_1b_2} \log \mathbb{P}[\text{PHSP}|b_1b_2]\} \end{aligned}$$

Length/sex frequency data, and selectivity

This is the change from the 2013 model; , and it required more effort than actually incorporating the HSPs. As noted in the main text, selectivity in the Indonesian fishery appears to have varied substantially in some years between 2002 and 2014, with clear bumps and dips that cannot be explained merely by “overdispersion” at the level of independent 1cm length classes— the approach used in 2013, where the overdispersion was estimated from a “model-free” pre-analysis. Ignoring these selectivity shifts, or treating them just as overdispersion, would tend to overweight the LSfreq data in the overall log-likelihood, potentially compromising the fit to the other datasets (POPs, HSPs, A@LS). The new model incorporates instead a year-specific random-effect spline which . The recorded LSfreq data $n_{\ell sy}^{\text{LS}}$ is currently represented as Poisson count data with:

$$\begin{aligned} \log \mathbb{E} [n_{\ell sy}^{\text{LS}}] = & \text{offset}(\log n_{sy\ell}) + \beta_y^{\text{LS}} + \text{tresid}_{s\ell} + \sigma_0(s, \ell) + \sigma_y(\ell) \quad (0.2) \\ n_{sy\ell} = & \sum_{a=8}^{A_{\max}} n_{sya} \mathbb{P}[\ell|sa] \end{aligned}$$

where β^{LS} is an intercept, $n_{sy\ell}$ and $\mathbb{P}[\ell|sa]$ come from the population dynamics, σ_0 is an overall selectivity following a prescribed functional form with estimable fixed-effect coefficients, and $\sigma_y(\ell)$ is the annual random-effect curves. The latter are chosen to be Duchon splines (Wood, 2017), which can be set up to penalize first-derivative penalties so that their default “preference”, in the absence of any data, is to be constant across length; better-known choices such as cubic splines will tolerate any linear trend without penalty, which allows too much freedom. The term $\sigma_0(s, \ell)$ can be specified in any reasonable way; options considered so far are constant (leading to asymptotic selectivity driven by residence time only, as in the 2013 model), and continuous-piecewise-linear with kinks at 165cm and 190cm. The latter allows dome-shaped selectivity, as described in the main text.

The ADT software currently only permits one random-effect variance to be estimated internally inside a model, and that has been used up here for recruitment variability. Consequently (and as with LSfreq overdispersion in 2013), the spline variability has to be pre-estimated. This was done by comparing LSfreqs (aggregated across sexes) in adjacent years; population-dynamics-driven changes should be slow enough that the main difference across a single year would be due to selectivity shifts. To avoid

having to specify any model for underlying length-frequency in the preliminary analysis, we fit a Gaussian GAM with Duchon splines to differences between successive annual *proportion-at-length* (i.e. normalized by total LSfreq sample size in each year), with approximate weights computed by the Delta-method to account for Poisson variability. The spline variance from this model should transfer directly to equation (0.2). The preliminary analysis is fitted twice, starting first from an odd-numbered year and then from an even-numbered year, and the two estimated spline variances are averaged.

Because true variability in population length frequency is ignored, this type of preliminary analysis will tend to *slightly* underweight LSfreq data in the main model (although, as noted in the main text, the LSfreq data actually still seem to be getting too much weight, and the model for sex frequency is clearly not adequate yet— there is some more work to do). Nevertheless, for a species like SBT with fairly slow turnover, it should be a simple and general way to allow for “uninteresting but unignorable” annual shifts in selectivity.

Age-at-length-and-sex data

Since selectivity is assumed to depend on length but not on age, each age-at-length-and-sex datum can be treated as an independent size-1 multinomial variable, with probabilities given by

$$\mathbb{P}[a|ls] = \frac{n_{sya} \times \mathbb{P}[\ell|sa]}{\sum_{a'} n_{sya'} \times \mathbb{P}[\ell|sa']}$$

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Appendix 2a: CSIRO project staff

The following CSIRO staff constituted the formal project team for this FRDC Project.

Name	Position	Organisation
Campbell Davies	Principal Investigator	CSIRO Oceans and Atmosphere
Mark Bravington	Senior Statistician	CSIRO Data61
Peter Grewe	Senior Geneticist	CSIRO Oceans and Atmosphere
Paige Eveson	Senior Experimental Scientist	CSIRO Oceans and Atmosphere
Jordon Aulich	Genetics Technician	CSIRO Oceans and Atmosphere
Peta Hill*	Genetics Technician	CSIRO Oceans and Atmosphere
Matt Lansdell	Research Technician	CSIRO Oceans and Atmosphere

*Currently PhD candidate at University of Tasmania

Appendix 2b: CSIRO staff - related projects

The following staff made important contributions to the delivery of the outputs and outcomes for this FRDC project through their roles in the listed related projects.

Name	Position	Organisation
Jessica Farley	Senior Experimental Scientist	CSIRO Oceans and Atmosphere
Craig Proctor	Senior Experimental Scientist	CSIRO Oceans and Atmosphere
Richard Hillary	Senior Fisheries Modeller	CSIRO Oceans and Atmosphere
Ann Preece	Senior Fisheries Modeller	CSIRO Oceans and Atmosphere
Pierre Feutry	Population Geneticist	CSIRO Oceans and Atmosphere
Paavo Jumppanen	Scientific Programmer	CSIRO Oceans and Atmosphere

Appendix 2c: Supporting agencies and companies

The following agencies and companies made the delivery of this project possible through their contributions to the sampling programs and support and advocacy for the work.

Agency/Company	Role
Research Institute for Tuna Fisheries, Ministry for Marine Affairs and Fisheries, Indonesia	Tissue, length and otolith sampling of adults through Long-term monitoring of SBT
Australian Southern Bluefin Tuna Industry Association (ASBTIA)	Advocacy for CKMR and support for sampling of juveniles in Port Lincoln
Protec Marine	Sampling of juveniles in Port Lincoln
Seatec	Sampling of juveniles in Port Lincoln
Commission for the Conservation of Southern Bluefin Tuna (CCSBT)	Funding for continue collection, archiving and sequencing of samples
Fisheries and Climate, ABARES	Support for CKMR in CCSBT
SBT fisheries management, SBTMAC and AFMA	Support for CKMR in domestically and in CCSBT
International Fisheries Section, DAWR	Advocacy for CKMR in CCSBT and direct co-investment in this FRDC project

Appendix 3: Related Projects

Next Generation Sequencing methods for Species, Provenance and Individual ID. CSIRO. Grewe, P.M. and Davies, C.R., 2012-2015. This strategic CSIRO appropriation project funded exploration of a range of Genotyping-by-sequencing platforms and marker types and the development of CKMR markers using these techniques for tuna, as well as provenance and species ID markers. *Where to now with Close-kin for SBT?* Bravington, M.V. and Davies, C.R., 2013. CCSBT-CSIRO. This project provided a strategic review of potential uses of CKMR into the future, including its use in the Operating model, as an independent time-series and as a potential input into future management procedures.

Close-Kin Mark-Recapture for SBT: options for the longer term. Bravington, M.V. 2014. CCSBT-CSIRO. This project completed a cost-benefit analysis for CCSBT to select between the existing

microsatellite markers vs adopting the new SNIp-based sequencing approach. The CCSBT agreed to proceed with the SNIp-based approach.

Estimating abundance, mortality and selectivity using Close-kin pairs. Bravington, M.V. et al., 2015. CSIRO. This project refined the theory and developed the abundance and mortality estimation model used to deliver the outputs of this FRDC project.

Collection and genotyping of 2015-2016 SBT samples for close-kin. Farley J. et al., 2015. CCSBT-CSIRO. This project covered the collection of biological samples (tissues, otoliths) required for CKMR in 2015 and 2016.

Fishery-independent estimate of spawning biomass of Southern Bluefin Tuna through identification of close-kin using genetic markers. Bravington M.V., Grewe P.G., Davies C.R. 2014. FRDC 2007/034. FRDC-CSIRO. The original application of CKMR approach to SBT. A subset of the samples and DNA extractions and associated data collected through this project will be used in this current FRDC project.