



Update on the SBT close-kin tissue sampling, processing and kin-finding 2025

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Abstract

The close-kin project is an ongoing monitoring program that provides data on the adult component of the population for use in the Cape Town management procedure and stock assessment models.

Muscle tissue samples collected from harvested juvenile southern bluefin tuna (SBT) in Australia in 2023 were subsampled, and DNA was extracted and sequenced. The kin-finding analysis to identify parent-offspring pairs (POPs) and half-sibling pairs (HSPs) was updated to include the new juvenile data. The results were provided to the CCSBT in May 2025. A total of 125 POPs and 221 high-confidence HSPs have now been identified, with a false negative rate for the HSPs of 0.32. Note the number of confidently identified HSPs has declined compared to last year's analysis (Farley et al. 2024), and correspondingly, the false negative rate has increased. This is because the overlap between the distributions of HSPs and less related pairs has continued to increase with more comparisons being made, so in order to ensure a very low number of false positive HSPs it was necessary to adjust the cutoff for determining HSPs upwards.

Between September 2024 and May 2025 (2024/25 spawning season), muscle tissue samples were collected from 1,148 adult SBT in Indonesia. While below the usual target of 1,500 samples (and well below the 3,200 samples proposed for this season to compensate for previous years' low coverage), it represents a marked increase and improves coverage for the adult population in close-kin analyses.

The 2025 sampling of 16,000 juvenile SBT in Australia has been completed.

Introduction

In 2013, the Extended Scientific Committee (ESC) developed a Scientific Research Plan (SRP) for southern bluefin tuna (SBT), identifying close-kin mark-recapture (CKMR) genetics as a high-priority activity for estimating adult abundance. Since 2014/15, the CCSBT has funded the collection and archiving of SBT muscle tissue, with DNA extraction and sequencing commencing in 2015/16. This work contributed to the completion of a second CKMR abundance estimation project that incorporated both parent-offspring pairs (POPs) and half-sibling pairs (HSPs), reported to the ESC in 2018 (Davies et al. 2018; 2020). Since 2018, the CCSBT has also funded the analysis of sequencing data to find POPs and HSPs (close-kin identification) on an annual basis, and this was formalised as an on-going monitoring activity under the 2023-2027 SRP in 2022. Table 1 summarises the work completed by spawning season since 2014/15. In 2019, the CCSBT endorsed an increase in genotyping targets from approximately 2,000 to 3,100 samples per year (combined adults and juveniles) to enhance POP-based cohort comparisons (Anon 2019). This paper provides an update on program activities in 2025.

Table 1. Summary of SBT close-kin work undertaken as part of CCSBT projects since 2015.

Sampling season	No samples for adults	No samples for juveniles	Genotyping completed	Kin-finding completed
2014/15	1500	1600	Yes	Yes
2015/16	1500	1600	Yes	Yes
2016/17	1500	1600	Yes	Yes
2017/18	1500	1600	Yes	Yes
2018/19	1500	1600	Yes	Yes
2019/20	1500	1600	Yes	Yes
2020/21	1500	1600	Yes	Yes
2021/22	0	1600	Yes	Yes
2022/23	148	1600	Juveniles complete Adults not complete	Juveniles only
2023/24	236	1600	Planned for 2025	Planned for 2026
2024/25	1148	1600	Planned for 2026	Planned for 2027

Muscle tissue collection

A total of 1,148 muscle tissue samples were collected from adult SBT at the Benoa Fishing Port in Indonesia between September 2024 and May 2025 (2024/25 spawning season). This is slightly lower than the target 1,500 samples but substantially higher than had been collected annually in the preceding three seasons (Table 1). Sampled fish ranged in size from 128 to 203 cm fork length (cm).

The target number of samples from juveniles was met in 2025 with 1600 samples collected at the tuna processors during harvest operations in Port Lincoln, South Australia, in April-July. Tissue was obtained from fish ranging from 89-105 cm fork length (FL) to cover the size range of 3-year-olds (see Preece et al. 2024). The tissue samples were placed in 2ml tubes and frozen according to protocols provided by CSIRO and will be transported to Hobart. The frozen muscle tissue samples are stored in consecutively labelled boxes with 100 positions (10 by 10) in each box (A01 through J10). Individual samples are given a unique identification label (e.g., SbPL2024_Bx01_A01).

DNA extraction and sequencing

Muscle tissue samples from juveniles collected in Port Lincoln (Australia) in 2023 were subsampled and DNA extracted. The DNA was extracted using a magnetic bead-based extraction protocol (Machery Nagel Nucleomag) kit on an Eppendorf EP motion robot to produce a 90uL archive and 30uL working stock of DNA in micro-titre format plates. Working stock plates of extracted DNA were shipped to Diversity Arrays Technology (DART) in Canberra for sequencing (referred to as “DARTcap”) of approximately 2000 single nucleotide polymorphic loci (SNPs). Archive plates of extracted DNA are stored in a dedicated -80°C freezer located at CSIRO Hobart. All sequencing

data were sent to CSIRO Hobart in early 2025 for inclusion in the kin-finding (below). There were no new adult data because very few adults were sampled in 2024/25 in Indonesia and the samples are still in Benoa. These samples will be sequenced and included in kin-finding as soon as they are transported to Australia and available for analysis.

Kin-finding

The kin-finding analysis database used for identification of POPs and HSPs was updated to include the DArT sequencing data for juvenile samples collected in 2023.

Prior to kin-finding, the sequencing data are used to “call the genotype” for each fish and locus in the data (i.e., to infer the pair of alleles present). This genotype-calling entails complicated algorithms developed by CSIRO specifically for DArTcap sequencing data and estimates the genotyping error rates for each locus, which is important in the identification of HSPs. A plate-level standardization was applied to the sequence count data from all years before calling the genotypes (Farley et al. 2019). This ensured that, for a given locus, the average count across all samples on a plate was the same for each plate.

A series of quality control (QC) steps were applied to the genotyped data to remove fish with unreliable genotype calls. These include:

- a test for heterogeneity to remove fish with an unexpectedly high number of heterozygous loci, which could be an indication of cross-contamination of DNA between individuals;
- a test of whether an individual's genotype could plausibly have been drawn from the species' allele frequencies as represented by the rest of the samples to remove fish potentially mis-identified as SBT; and
- a test for an over-representation of null alleles in each individual genotype to remove degraded samples.

After applying the QC steps, 11,259 adults and 20,476 juveniles remained for kin-finding (Table 2). Note that this table only includes samples that were sequenced using DArTcap and analysed in this report; it does not include samples that were sequenced prior to 2015 using microsatellite markers (recall that the genotyping method changed after 2015 from using microsatellites to DArTcap sequencing; see Bravington et al. 2015; 2017).

POP-finding

We used the genotype data to identify POPs using the same method as previous years, which is a modified Mendelian-exclusion statistic referred to as the Weighted-PSeudo-EXclusion (WPSEX) statistic (see Appendix B of Bravington et al. 2017).

Figure 2 shows the histogram of the WPSEX statistic comparing across all possible genotyped adult-juvenile pairs (20,476 juveniles x 11,259 adults = 230.5 million comparisons). The POPs are visible as a small bump to the left of the dashed blue line. Most of the histogram (to the right) has been truncated, because otherwise the POPs are too few compared to the gigantic bump of

unrelated pairs (the peak of which is around 0.116, where theory predicts it should be, based on allele frequencies of each locus) and could not be visualized. The giant bump of unrelated pairs drops off quickly to the left of ~ 0.075 , and the “tail” between ~ 0.045 - 0.075 will contain a number of adult/juvenile HSPs and grandparent-grandchild pairs. The gap between this “tail” and the POPs bump has become less distinct over the years, which is to be expected given the number of comparisons has increased exponentially. In future, additional analyses may be required to help determine whether pairs in this uncertain zone between ~ 0.04 - 0.05 are POPs or less related pairs.

The number of POPs identified in this data set is 82. Including the POPs that were identified previously using microsatellites, we now have a total of 125 pairs. The breakdown by juvenile birth year and adult capture year is given in Table 3.

Table 2. Number of fish used in the kin-finding analyses this year after quality control checks were applied. For the adults, samples were collected from Indonesia in the fishing season ending in the year shown (i.e., samples collected over the 2005/06 fishing season are referred to as year 2006).

Year	Adults	Juveniles
2006	0	1317
2007	0	1325
2008	0	1356
2009	0	1347
2010	972	1315
2011	958	963
2012	536	876
2013	959	903
2014	922	899
2015	0	953
2016	951	854
2017	969	948
2018	700	756
2019	1440	1449
2020	1421	1512
2021	1431	1384
2022	0	1176
2023	0	1143
Total	11259	20476

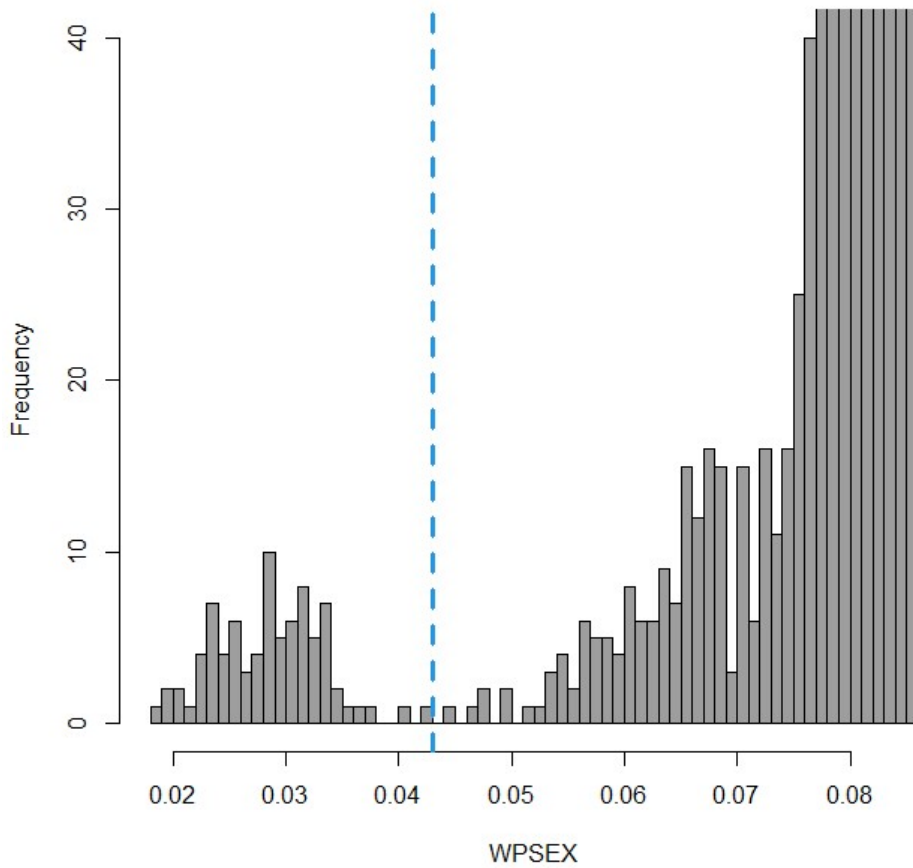


Figure 2. Histogram of the weighted-pseudo-exclusion (WPSEX) statistic for identifying parent-offspring pairs (POPs). Low values (below the vertical blue dashed line) indicate POPs. The x-axis is right-truncated to omit the large peak of unrelated pairs to the right.

Table 3. Number of parent-offspring pairs (POPs) (including those identified using microsatellites and DArTcap data) broken down by juvenile birth year (rows) and adult capture year (columns). NA indicates that no POPs were possible either because no samples exist for that combination of years, or the adult capture year is before the juvenile birth year.

		Adult recapture year															
		2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021
Juvenile birth year	2002	0	0	0	0	0											
	2003	0	5	1	2	0	0	0	1	0		1	0	0	0	0	0
	2004	0	2	0	0	3	0	0	0	0		0	0	0	0	1	0
	2005	1	4	5	4	1	0	0	1	2		0	0	0	0	0	0
	2006		4	3	2	0	0	0	0	0		0	0	0	0	0	2
	2007			3	4	1	3	2	0	2		0	1	0	0	1	1
	2008					0	1	1	1	0		0	0	2	0	0	0
	2009					0	1	1	1	0		0	0	0	1	1	0
	2010					0	0	1	4	0		2	0	0	1	0	0
	2011						0	1	2	1		2	0	0	0	0	0
	2012							0	1	1		0	0	1	0	0	1
	2013								0	0		1	1	3	1	1	0
	2014									0		0	1	0	0	1	0
	2015											1	0	0	0	0	0
	2016											0	2	1	0	1	1
	2017												3	0	1	2	0
	2018													1	2	3	0
	2019														1	0	2
	2020															1	1

HSP-finding

HSPs were again identified using a pseudo-log-odds-ratio (PLOD) statistic to measure the relative probability of a pair of fish having their observed genotypes if they are HSPs compared to if they are unrelated (see Appendix C of Bravington et al. 2017).

When using the PLOD statistic to compare all possible pairs of juveniles ($20,476 \times 20,476 = 419.3$ million comparisons), we do not get a clear separation between the distribution for HSPs and that for unrelated/less related fish, in particular half-thiatic pairs (HTPs, e.g. uncle and half nephew). This has been noted as a problem in the past several years, and the overlap keeps increasing as the number of comparisons increases. We dealt with this overlap issue using the same approach as in 2021 to 2023 (e.g. Farley et al. 2022), limiting the comparisons to juveniles born less than 9 years apart (or equivalently, sampled less than 9 years apart since all juveniles are age 3 when sampled). Since HTPs are likely to be greater apart in age than 9 years, this reduces the number of comparisons between fish that are potentially HTPs, thus reducing the size of the problematic HTP bump while not excluding too many potential HSPs. To address this issue long term, a high-quality genome assembly for SBT was developed in 2021 in collaboration with the Wellcome Sanger Institute (UK). New algorithms are being developed that leverage linkage information between genetic markers gained from the genome assembly to improve the accuracy of kin-finding. These algorithms have taken longer than anticipated to develop, in part because the approach has morphed over time. Good progress has been made in the past year, but the algorithms still require further testing before applying to SBT.

The PLOD statistic for comparisons between juveniles born less than 9 years apart is shown in Figure 3. The four pairs with PLOD values to the right of 150 are full sibling pairs (FSPs)¹. The division between HSPs and less related pairs is still not as clear as we would like however, setting a PLOD cut-off value of 55 for HSPs (increased from last year when it was 50; Farley et al. 2024) results in only 3 expected false positive HSPs and 221 pairs that we are quite confident are HSPs. Note that we used the theoretical means and approximate variances of the PLOD distributions for HSPs and unrelated/less related pairs (calculated as part of the kin-finding software) to determine the expected number of false positives. The breakdown in numbers of identified HSPs by birth year is given in Table 4.

An inevitable consequence of ensuring that false positives are rare is that a reasonable number of false negatives will be present. Using the expected PLOD distribution for HSPs, we estimated the true number of HSPs to be about 32% higher than 221 because of false negatives. The false-negative rate is allowed for in the population modelling, so although the increasing value is not ideal, it can be accounted for within the model as long as we have a good estimate of it (Bravington et al. 2017).

¹ Note that all four FSPs were within-cohort pairs, as one would expect for a large adult stock.

Table 4. Number of half-sibling pairs (HSPs) broken down by birth year of younger sibling (rows) and older sibling (columns). Note that comparisons were only made between juveniles born less than 9 years apart.

	Juvenile birth year	Adult recapture year																	
		2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020
	2003	2	4	2	1	0	0	1	0	0									
	2004		5	2	6	2	2	1	0	0	2								
	2005			5	3	3	3	0	4	1	0	0							
	2006				8	3	1	3	4	3	0	1	1						
	2007					3	3	2	1	2	2	2	1	2					
	2008						5	1	0	2	3	0	1	0	2				
	2009							1	1	1	0	0	0	0	4	2			
	2010								2	1	2	1	0	1	1	0	0		
	2011									3	2	1	0	2	4	1	0	0	
	2012										2	2	1	1	1	2	1	0	0
	2013											2	4	0	0	1	0	0	0
	2014												2	2	1	4	3	1	0
	2015													4	2	1	3	0	0
	2016														6	3	4	6	1
	2017															5	2	3	4
	2018																5	2	0
	2019																	4	1

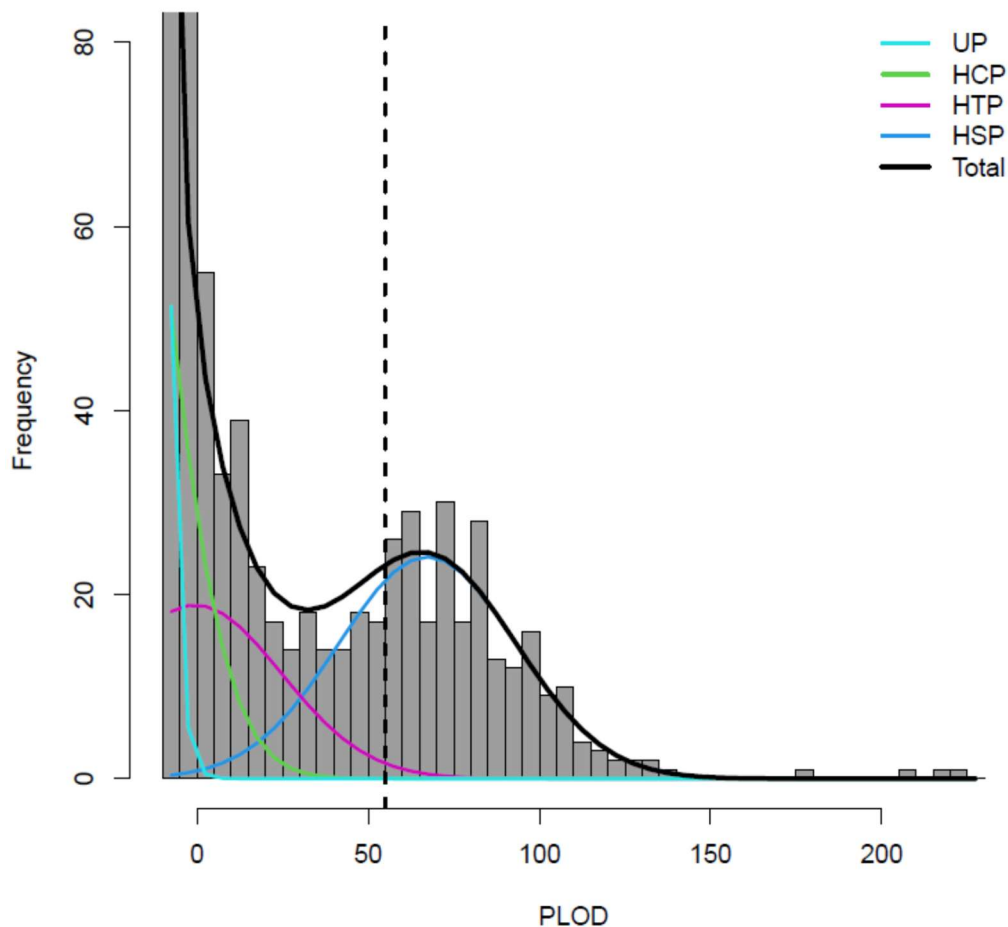


Figure 3. Histogram of the pseudo-log-odds-ratio (PLOD) statistic for pairwise comparisons of juveniles born less than 9 years apart. The approximate PLOD distributions for unrelated (UP), half-cousin (HCP), half-thiatic (HTP) and half-sibling (HSP) pairs are shown. With a lower PLOD cut-off value of 55 for HSPs, we expect ~3 false-positive HTPs. Note that the x-axis is left-truncated to omit the gigantic peak of UPs to the left.

Summary

This year we successfully completed:

- 1) The 2025 tissue sampling in Australia of 1,600 juveniles
- 2) The 2024/25 tissue sampling in Indonesia of 1,148 adults.
- 3) Tissue subsampling and DNA extraction and sequencing of juvenile samples collected in Port Lincoln in 20243.
- 4) Kin-finding (POPs and HSPs) to include the 2023 juvenile samples.

An updated dataset of identified SBT POPs and HSPs was provided to the CCSBT in May 2025. To date, a total of 125 POPs and 221 high-confidence HSPs have been identified with the false negative rate for HSPs estimated to be 0.32.

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